

# **Function and Analysis of Post-translational Protein Modifications**

Vorlesung 4610

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# Posttranslational Modifications of Proteins

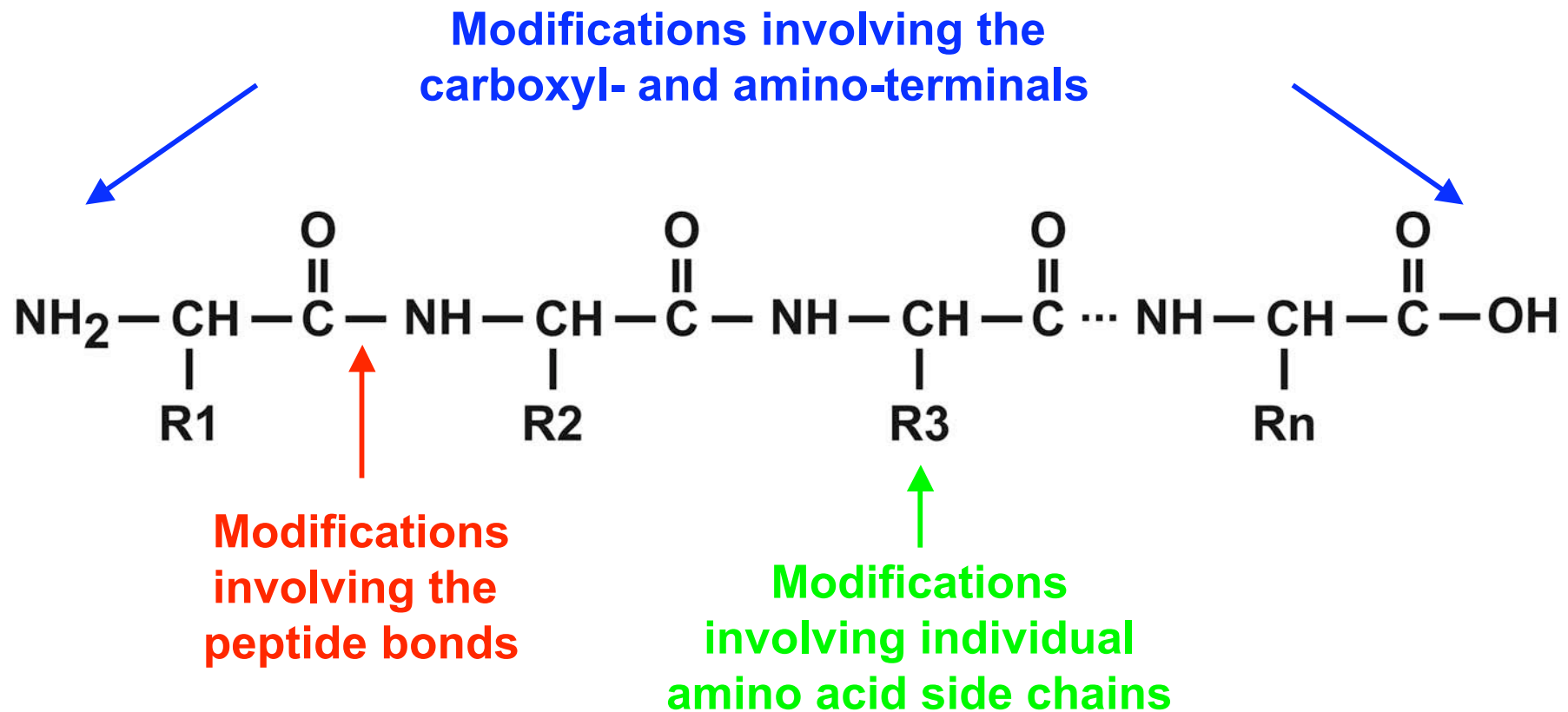
<http://www.abrf.org/index.cfm/dm.home?AvgMass=all>

Approx. naturally occurring 260  
PTMs listed!

# Posttranslational Modifications of Proteins

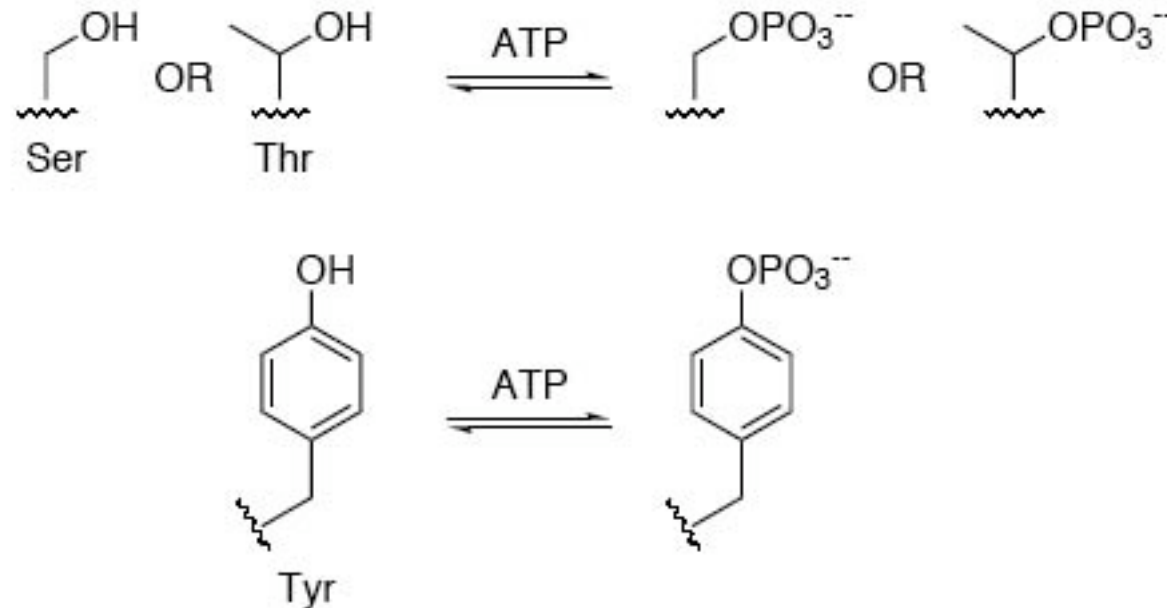
- Covalent attachment of chemical groups to a protein
- Some modifications occur while the protein is translated and exit from the ribosome
- Other modifications occur only after protein translation
- Require dedicated enzymatic catalysis

# Possible Sites of Modifications in Proteins



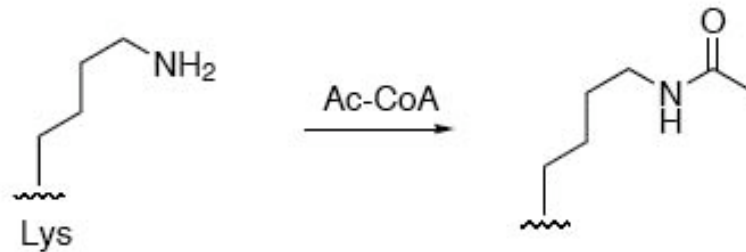
# Some of the Side Chains Enzymatically Modified by PTMs

- Ser, Thr, Tyr

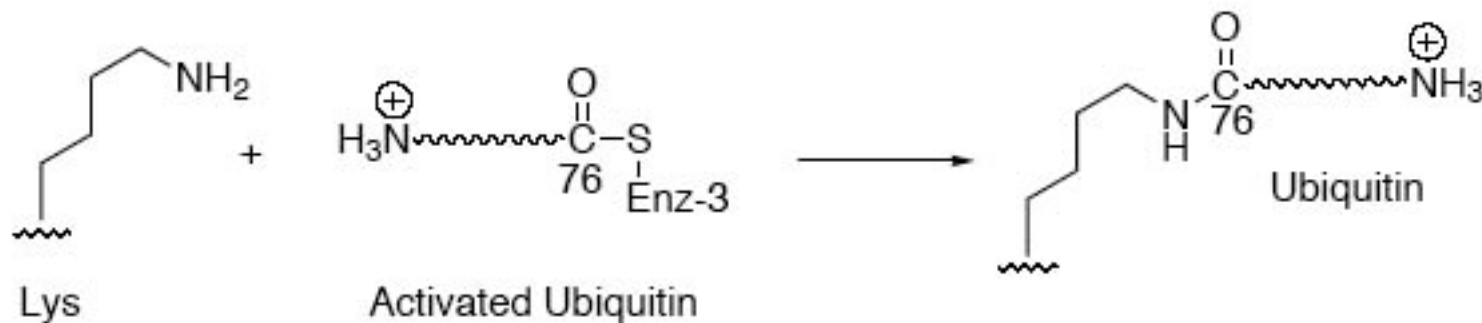


# Some of the Side Chains Enzymatically Modified by PTMs

- Lys acetylation

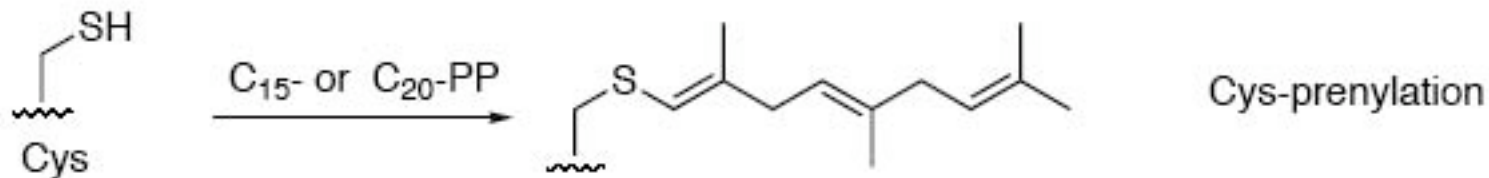
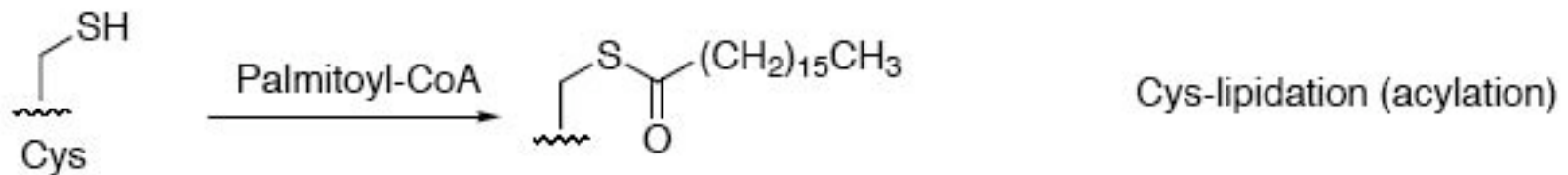


- Lys ubiquitination



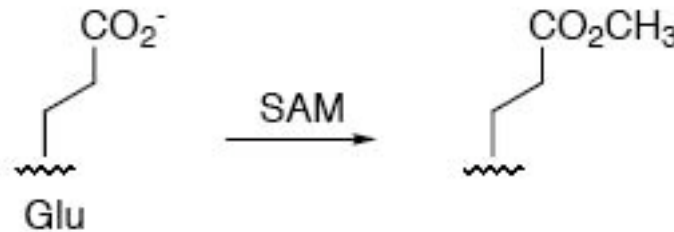
# Some of the Side Chains Enzymatically Modified by PTMs

- Cys lipidation
- Acylation by long chain fatty acyl CoAs ( $C_{14}$ ,  $C_{16}$ )
- Prenylation by  $C_{15}$  (farnesylation)

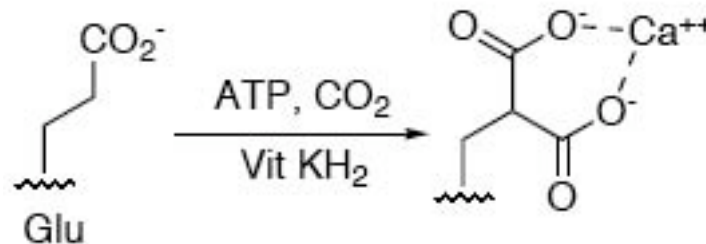


# Some of the Side Chains Enzymatically Modified by PTMs

- Glu methylation



- Glu carboxylation (blood coagulation cascades)



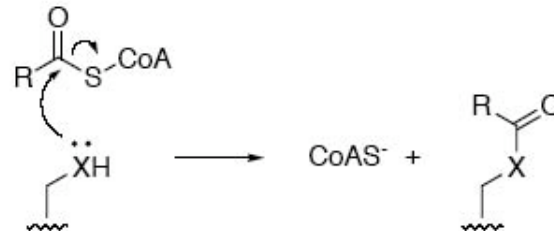


# General Reaction Scheme for PTM of Proteins

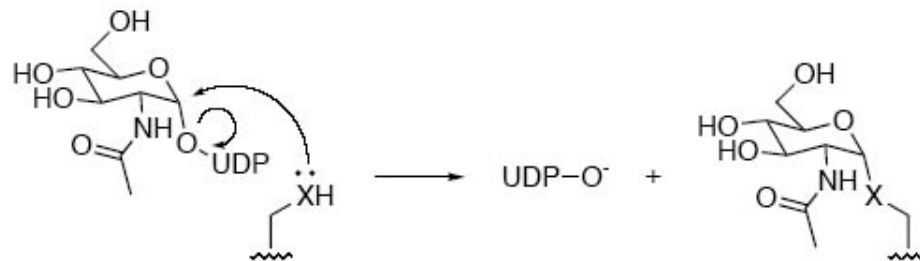
- ATP → phosphoproteins



- AcylCoAs → acylated lipoproteins

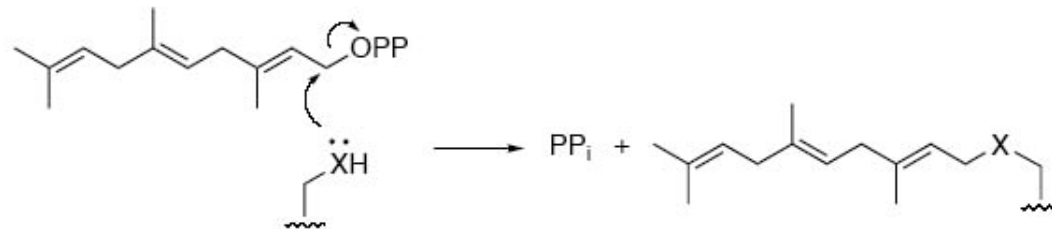


- UDPGlcNAc → glycoproteins

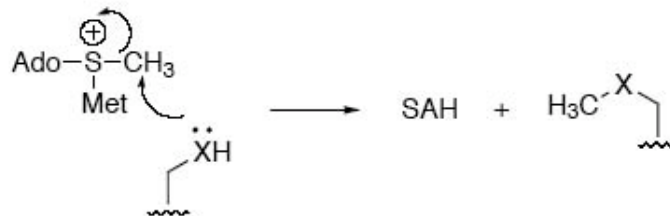


# General Reaction Scheme for PTM of Proteins

- Farnesyl-PP, Geranylgeranyl-PP → prenylated lipoproteins



- SAM → Methylated proteins



# Cellular Sites of Major Post-translational Modifications

## Site

**Cytoplasm**

## Modification

**Removal of initiating Met**

**Acetylation of N-terminus**

**Myristoylation of N-terminus**

**O-Glycosylation with GlcNac**

**Addition of palmitoyl groups**

# Cellular Sites of Major PTM's

## Site

## Modification

**Mitochondria/Chloroplasts**

**Cleavage of Signal Peptides**

**Golgi Apparatus**

**Modification of N-glycosyl groups,  
O-glycosylation with GalNAc**

**Secretory Vesicles/Granules**

**Amidation of C-terminus  
Proteolytic processing of some precursors**

# Cellular Sites of Major PTM's

## Site

## Modification

**ER**

**Cleavage of signal peptides**

**Core glycosylation of Asn residues**

**Addition of palmitoyl and glycosyl-phosphatidylinositol**

**Hydroxylation of Pro/Lys in procollagen**

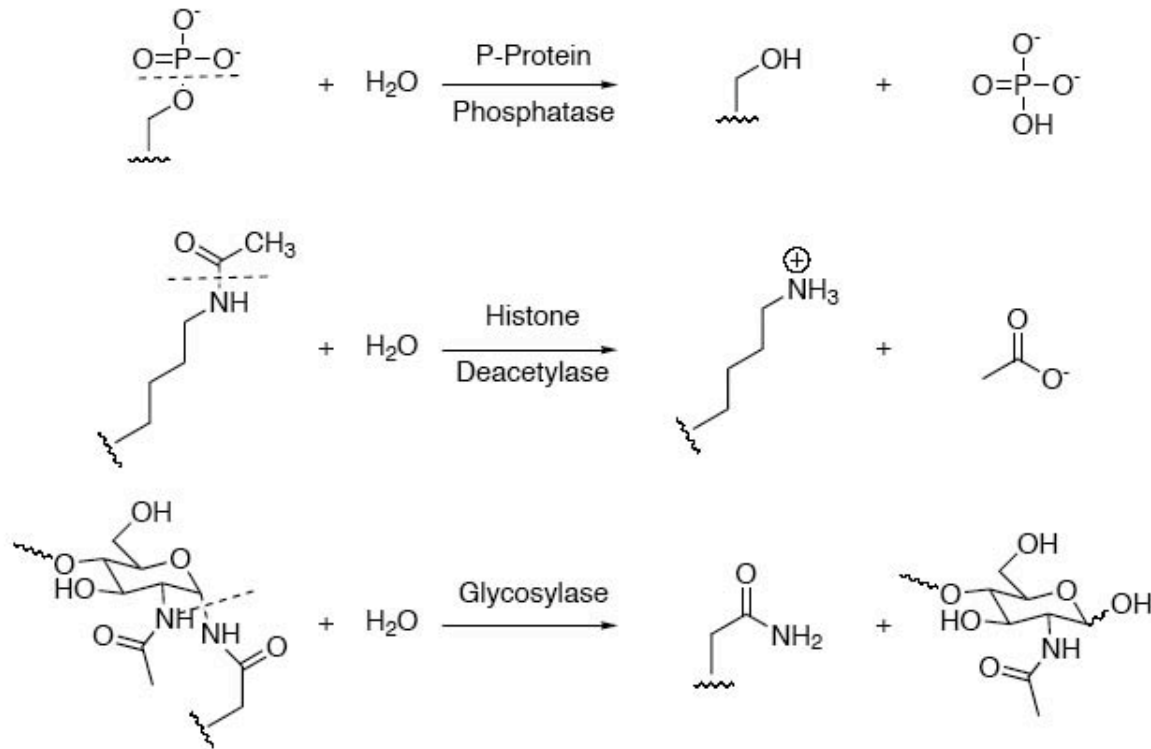
**Disulfide bond formation**

# Functions Enabled by Posttranslational Modification

- Alterations in local folding of proteins
  - E.g. transitions between unstructured and structured regions
  - Generation of charge pairs
- Marking proteins for degradation
  - Proteasome targeting
- Marking chromatin for transcriptional regulation
- Changing the intracellular or extracellular addresses of proteins
  - Signal peptides direct proteins to ...
  - Plasma membrane
  - Secretory pathway
  - Mitochondria
  - Cytosol
- Inactive apo to active holo forms of enzymes

# Sets of Enzymes

- PTMs are generally stable modifications
- Reversal of modification requires a distinct set of enzymes



# Modifications Involving the $\alpha$ -amino Group

- $N^{\alpha}$ -acylation
  - formyl
  - acetyl
  - pyruvoyl
  - $\alpha$ -ketobutyryl
  - glucuronyl
  - pyroglutamate
  - murein



# Formylation of the $\alpha$ -Amino Group

- N $^{\alpha}$ -formyl-Met
  - forms the start of the nascent protein chain in prokaryotes
  - Eukaryotic start signal is Met
- Deformylase may remove the CHO group
- Aminopeptidase later removes Met from some, but not all chains
- N $^{\alpha}$ -formyl-Gly
  - occurs in honey bee melittin
- Murein derivatives
  - link *E. coli* peptidoglycan and membrane lipoprotein

# Protein Acetylation

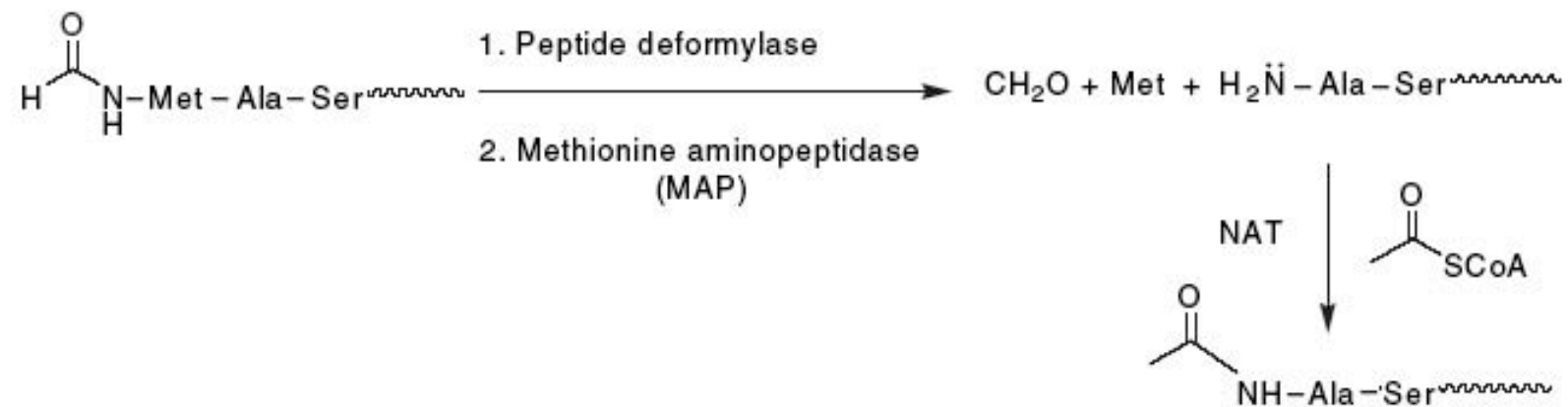
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# Protein N-Acetylation

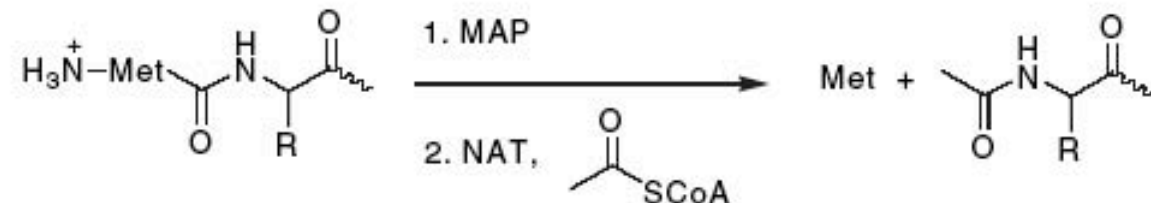
- In about 50% of yeast proteins and about 80-90% of higher eukaryotic proteins
- Very rare in *E. coli*
- S5 (N-Ac-Ala-Arg-...), S18 (N-Ac-Ala-His-...), L12 (N-Ac-Ser-Ile-...)

# Protein N-Acetylation

## A. Prokaryotes:



## B. Eukaryotes:



# Specificity of Protein N-Acetylation

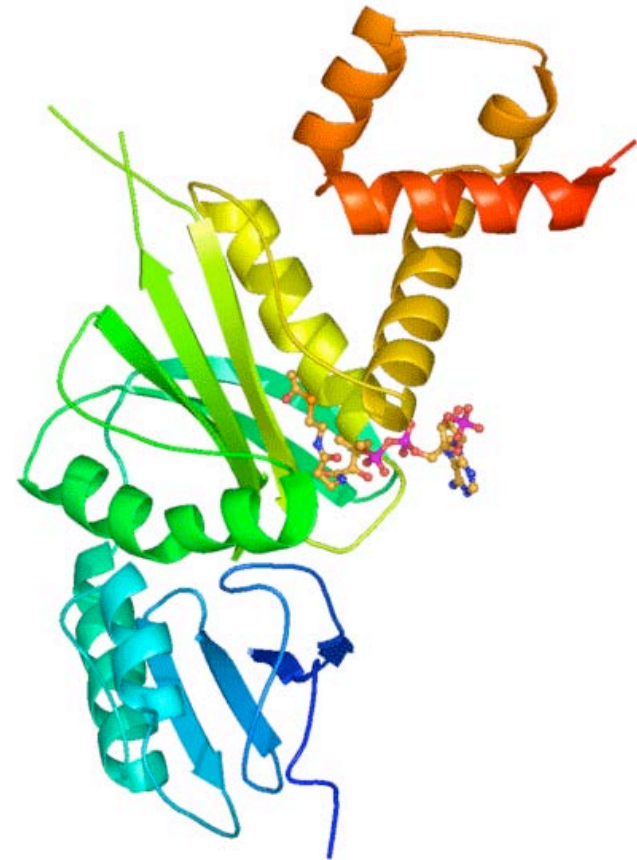
- Lies in the specificity of MAP
- MAP cleaves preferentially
  - Met-Gly
  - Met-Ala
  - Met-Ser
  - Met-Cys
  - Met-Thr
  - Met-Pro
  - Met-Val
- Uncovered Ser, Ala, Gly, Thr, get acetylated

# N-acetyl Transferases (NATs)

- In yeast, 3 NATs (NAT A, B, and C)
- NATB and NATC acetylate proteins with Met<sub>1</sub> still in place
- NATB recognises M-E, M-D, M-Q, M-M
- NATC recognises M-I, M-L, M-W, M-F
- NATA acetylates S, A, G, T

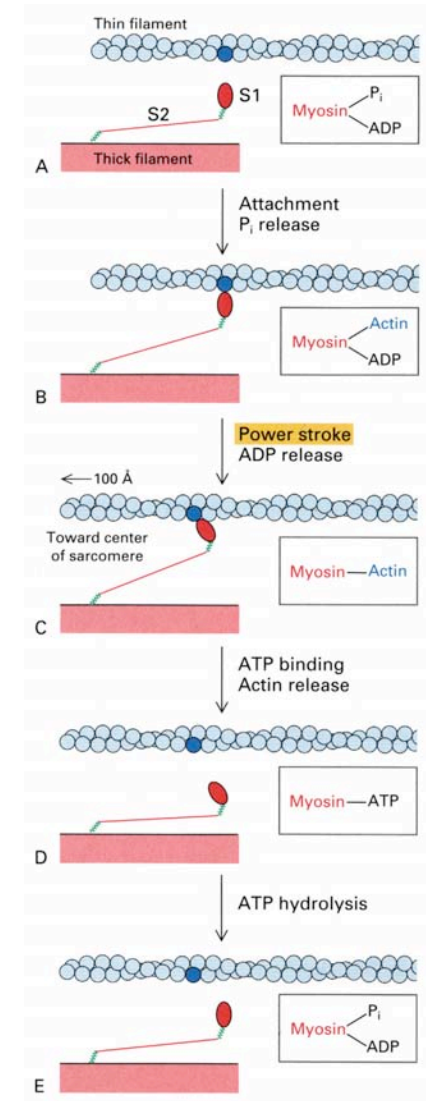
# N-acetyl Transferases

- In eukaryotes, hundreds of NATs exist
- Involved in K acetylation of histones
- Control
  - transcriptional activation
  - Chromatin assembly
  - DNA replication
  - Involved in N-acetylation of aminoglycoside antibiotics, results in decreased affinity of the drug for its target



# Biological Significance of N-terminal Acetylation

- Unclear in eukaryotes!
- Functions only detected on a case-by-case study
- Actins are known to be acetylated at N-terminus
  - N-terminus is M-E...
  - E exposed by aminopeptidases
  - Further acetylation to yield mature N-acetyl-E
  - Strengthens interaction between actin and myosin



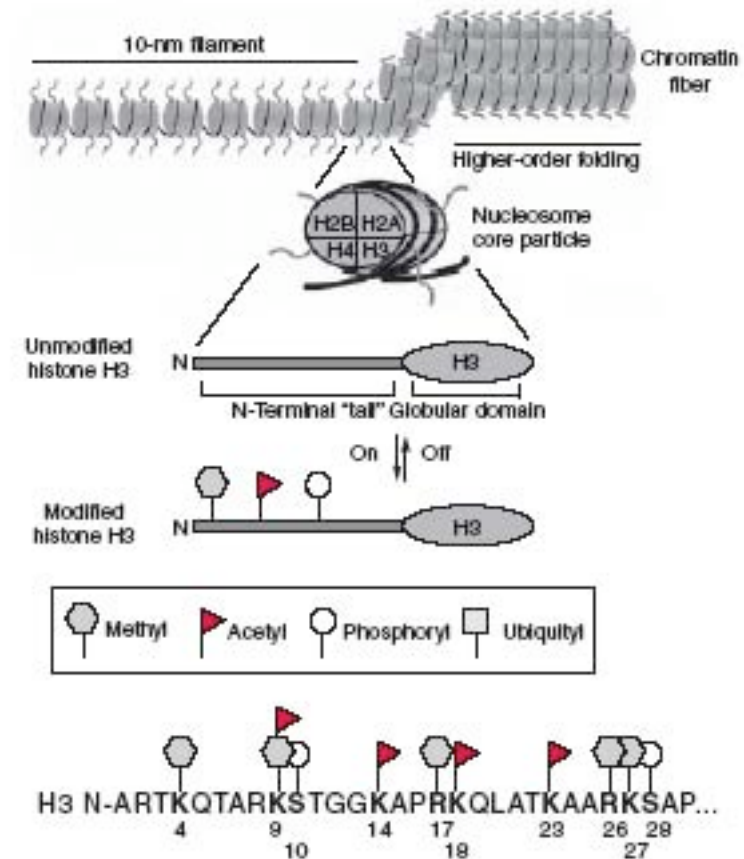
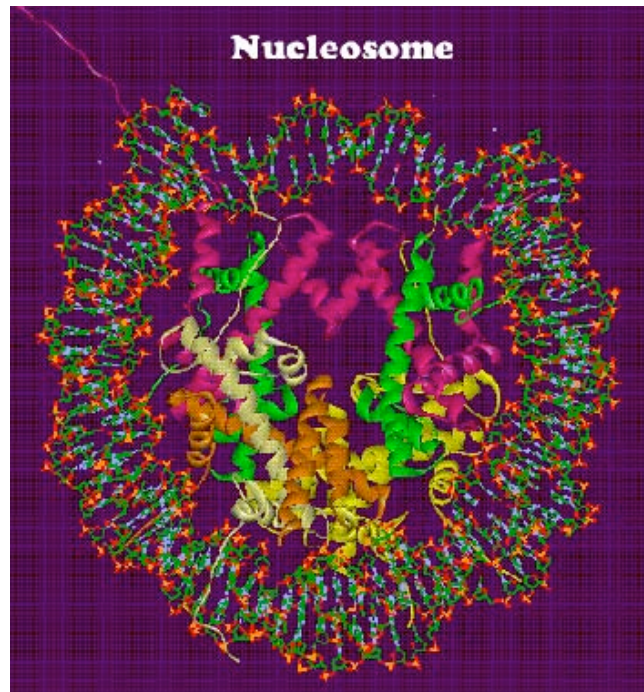


# N-Acetylation of Lysine- $\epsilon$ -NH<sub>2</sub> Side Chains

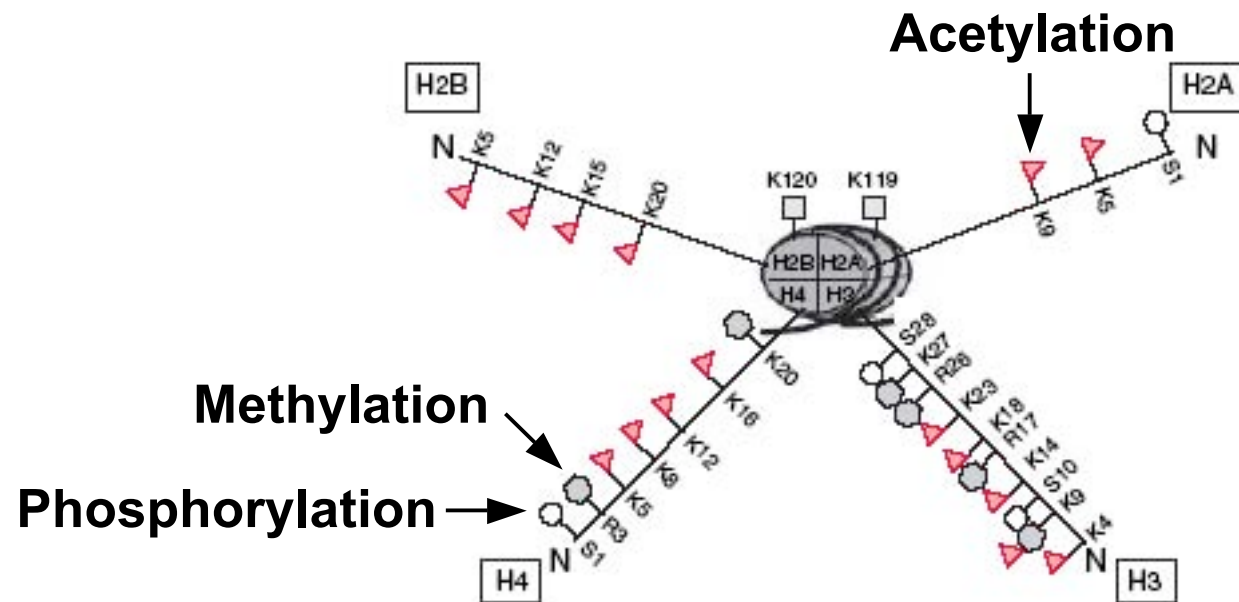
- Interest in protein N-acetylation is on regiospecific modification of K side chains
- Large number of PTMs detected on histones
- Transcriptional coactivators and corepressors turned out to be HATs and HDACs
- N-terminal regions of histones are flexible and amenable to PTMs

# N-Acetylation of Lysine- $\epsilon$ -NH<sub>2</sub> Side Chains

- Chromatin contains H2A, H2B, H3 and H4
- Histone core (H2A)<sub>2</sub>(H2B)<sub>2</sub>(H3)<sub>2</sub>(H4)<sub>2</sub>
- 145-147 bps around core



# Histone Acetylation

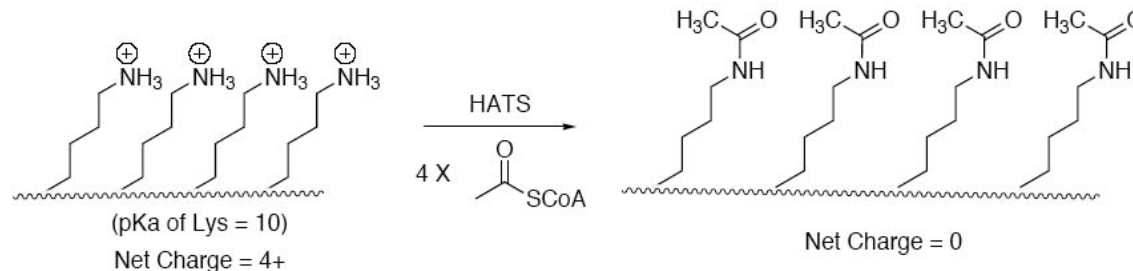


# Histone Acetylation

- MS analysis revealed acetylation on...
- H2A (K<sub>5</sub> and K<sub>9</sub>)
- H2B (K<sub>5</sub>, K<sub>12</sub>, K<sub>15</sub>, K<sub>20</sub>)
- H3 (K<sub>9</sub>, K<sub>14</sub>, K<sub>18</sub>, K<sub>23</sub>)
- H4 (K<sub>5</sub>, K<sub>8</sub>, K<sub>12</sub>, K<sub>16</sub>)
- For two copies each of histone: 28 potential acetylation sites
- Yeast: 13 acetylation sites found/octamer
- 50% posttranslational utilization
- Immense combinatorial possibilities








# Consequences of Histone Acetylation

- K's cationic at physiological pH
- N-acetylation quenches positive charges



- Electrostatic weakening of histone/DNA interactions
- Opening of the chromatin
- Allows TFs to bind to promoter regions

# Consequences of Histone Acetylation

H3 N-Termini	Modification	Function
	Unmodified	Silencing
	Acetylated	Transcription
	Acetylated	Histone deposition?
	Phosphorylated	Mitosis/Meiosis
	Phos/Acetyl	Transcription
	Methylated	Transcription?
	Higher-order combinations	?

# Histone Acetyltransferases - A Family

- Family of GNATs (Gcn5-related N-acetyltransferases)

yGcn5



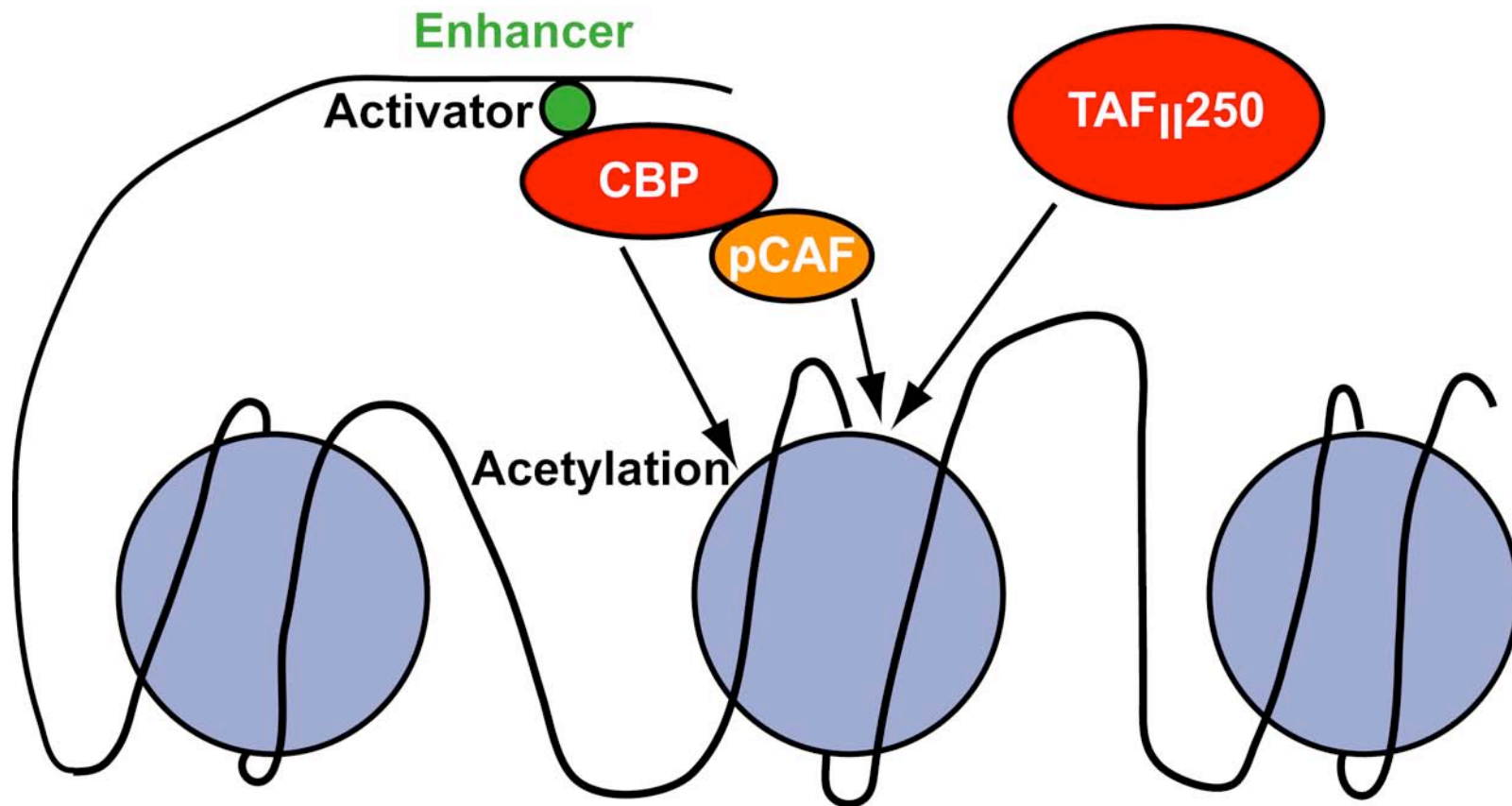
CBP



TAF<sub>II</sub>250

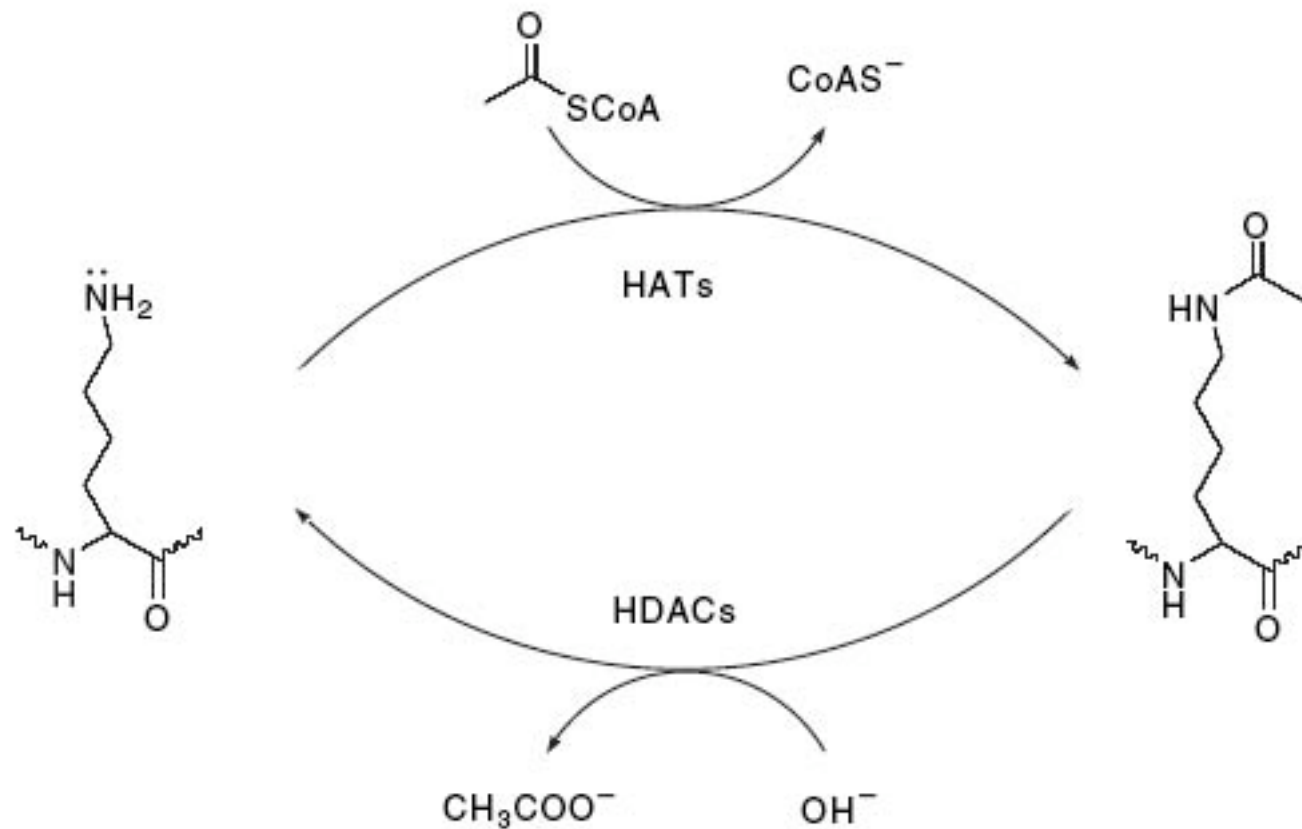


# Transcriptional Activation by HATs





# Histone Deacetylases (HDACs)

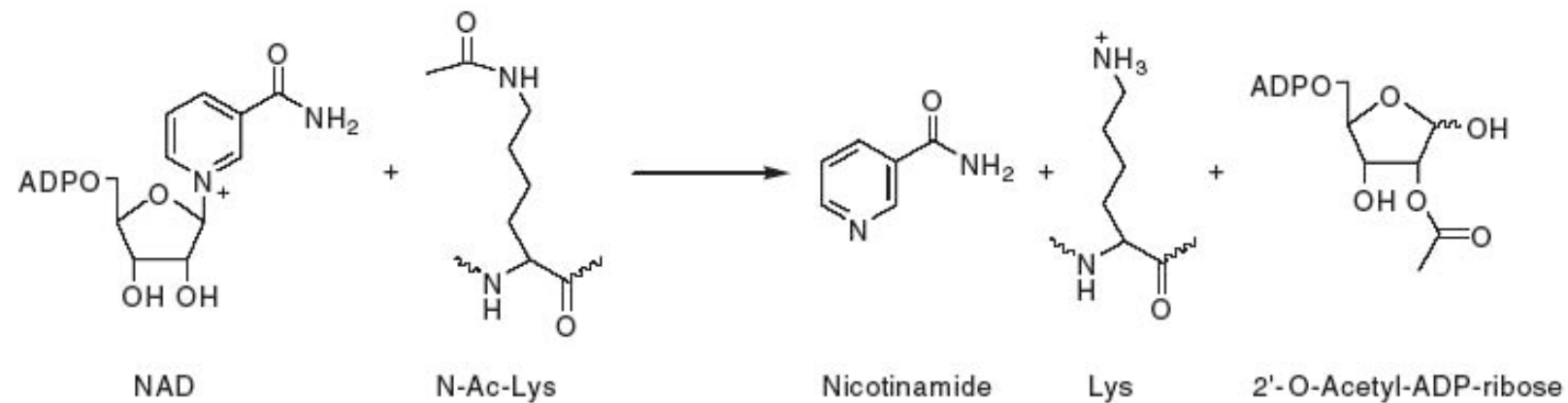


# Histone Deacetylases (HDACS)

- HDACS are corepressors of transcription
- Maintain histone tails in hypoacetylated state
- Leads to chromosome condensation
- Silence promoters
- 2 distinct families
  - HDACs that release the acetyl moiety as acetate
  - Sirtuins (silent information regulator)

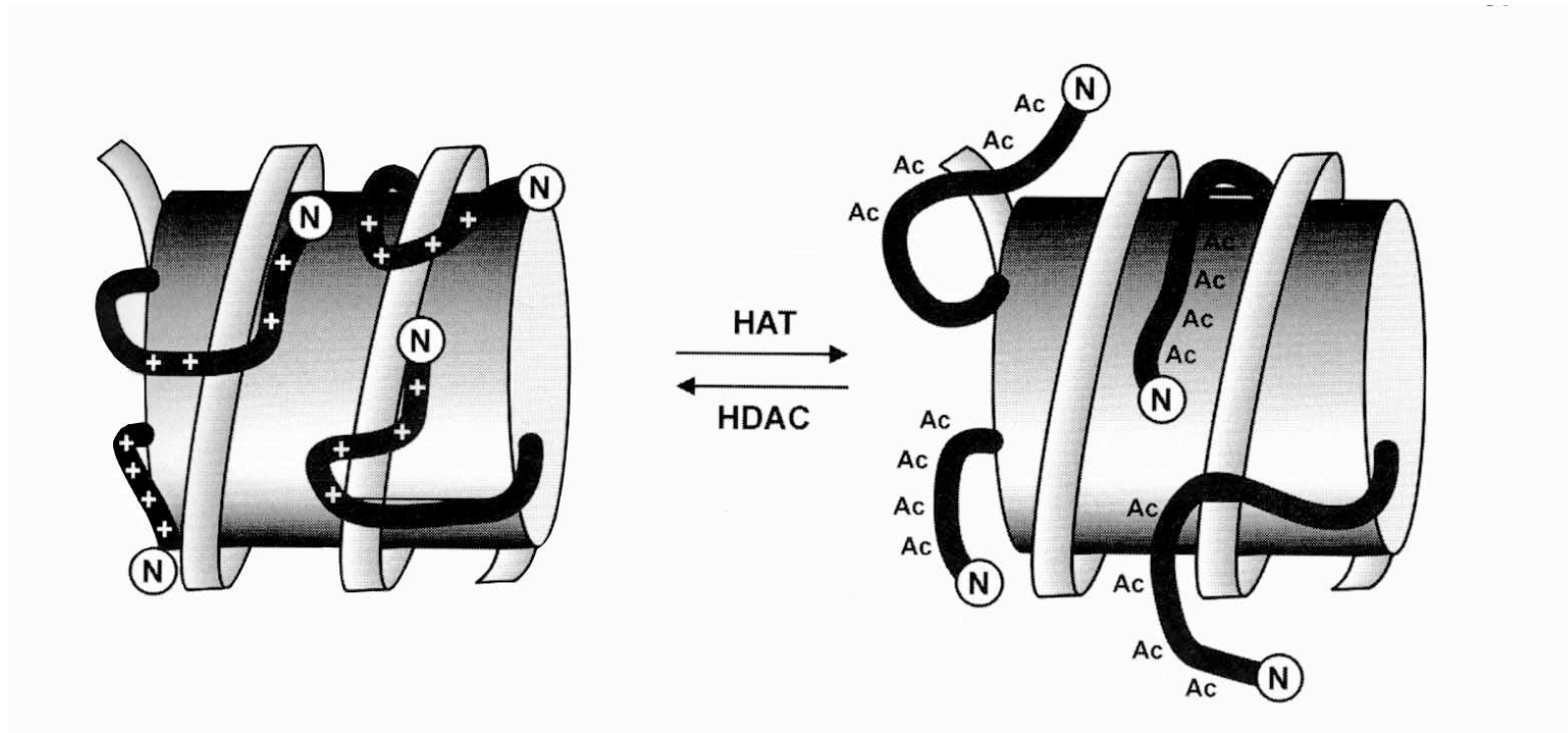
# Sirtuin HDACS

Sirtuin reaction stoichiometry:



Function and Analysis of Post-translational Protein  
Modifications

# Influence of Histone Acetylation and Deacetylation on Nucleosome Structure



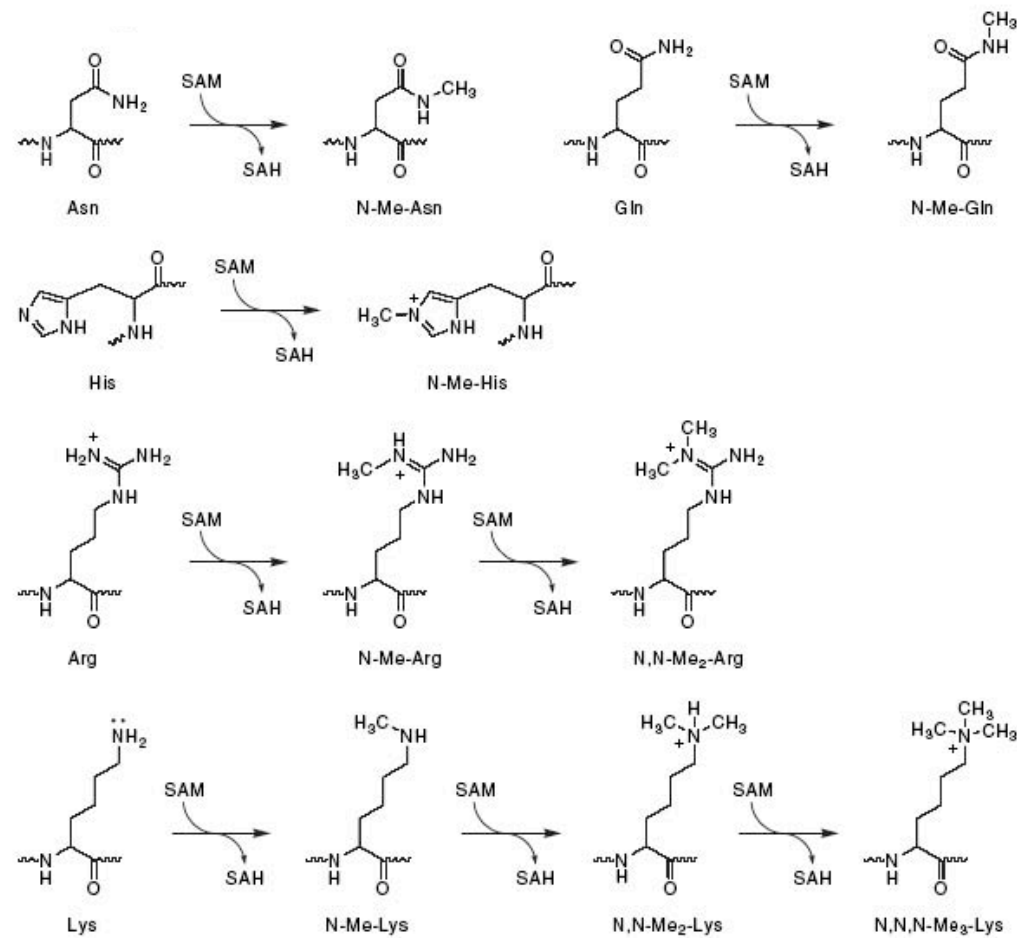
# Protein Methylation

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Modifications

# Protein Methylation

- Occurs on N- or O-atoms
- Methylation of  $\text{-COO}^-$  covers up negative charge
- N-methylation of Ks does not alter charge, increases hydrophobicity
- Di- and trimethylation of Ks increases both hydrophobicity and steric bulk
- Affects protein-protein interactions
- Occurs on  $\epsilon$ -amino group of K, imidazole ring of H, guanidino group of R, amides of Q and N
- N-methylation irreversible

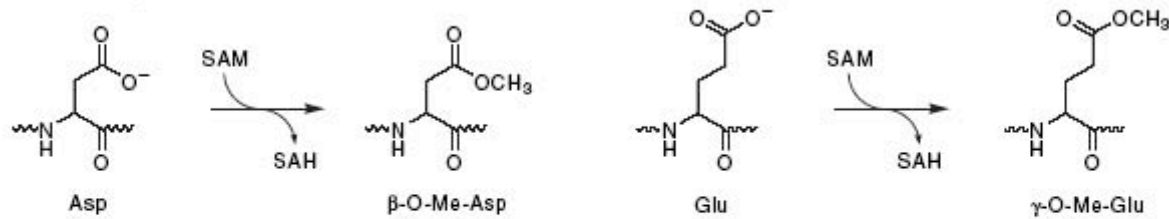
# N-Methylations



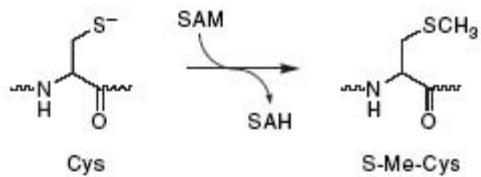
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# O-, S-, and C-Methylations

## B. O-Methylations:

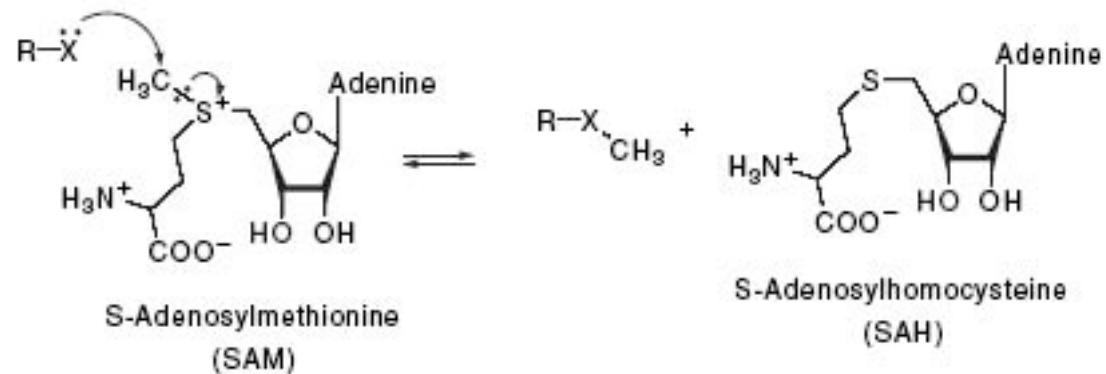
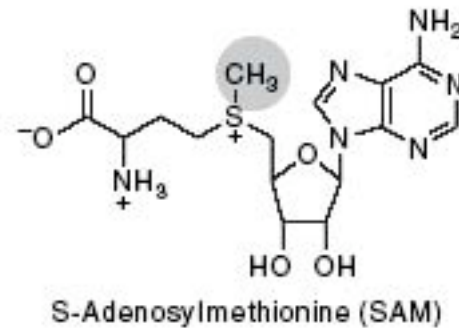


## C. S-Methylations:



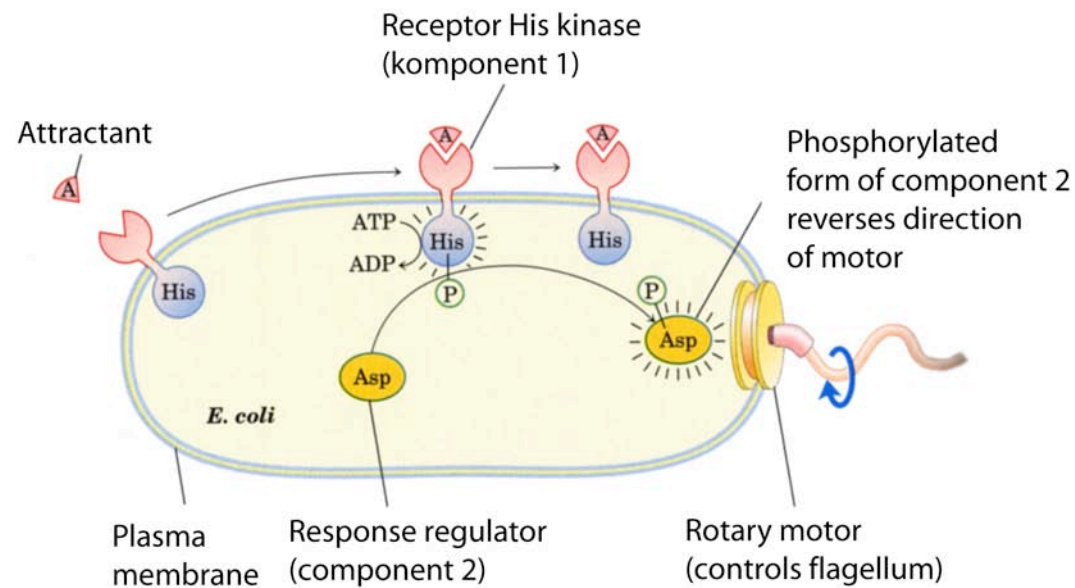


# One Carbon Donor



Function and Analysis of Post-translational Protein Modifications

# Protein O-Methylation Occurs in Bacterial Chemotaxis



# Protein O-Methylation

- Occurs in bacterial chemotaxis
- Transmembrane receptors for
  - Asp (Tar), Ser (Tsr), Peptides (Tap)

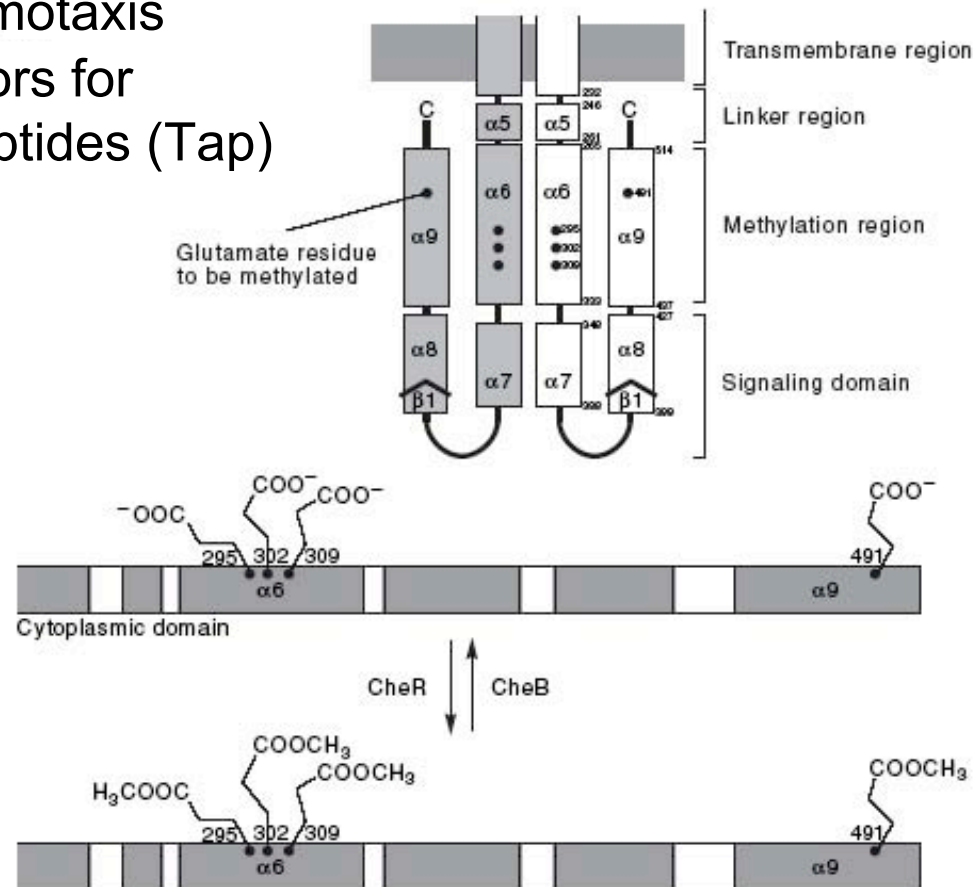


Diagram illustrating the structure of the chemotaxis receptor complex in *E. coli*, showing the interaction between the Maltose-binding protein (MBP) and the receptor, and the signaling pathway involving CheW, CheA, CheB, and CheC.

The diagram is divided into three regions: Periplasm, Membrane, and Cytoplasm.

**Periplasm:**

- Maltose-binding protein (MBP) is shown bound to the N-terminus of the receptor.
- Aspartate is shown as a ligand.
- Transmembrane helices TM1 and TM2 are labeled.
- Cytoplasmic helices  $\alpha 3$  and  $\alpha 4$  are labeled.

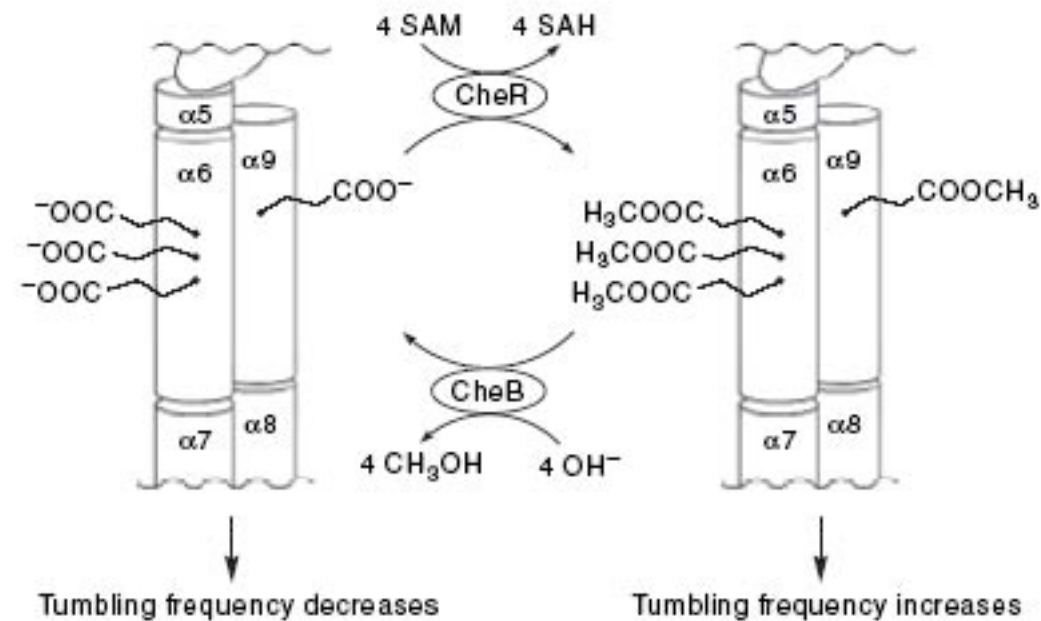
**Membrane:**

- The receptor spans the membrane.

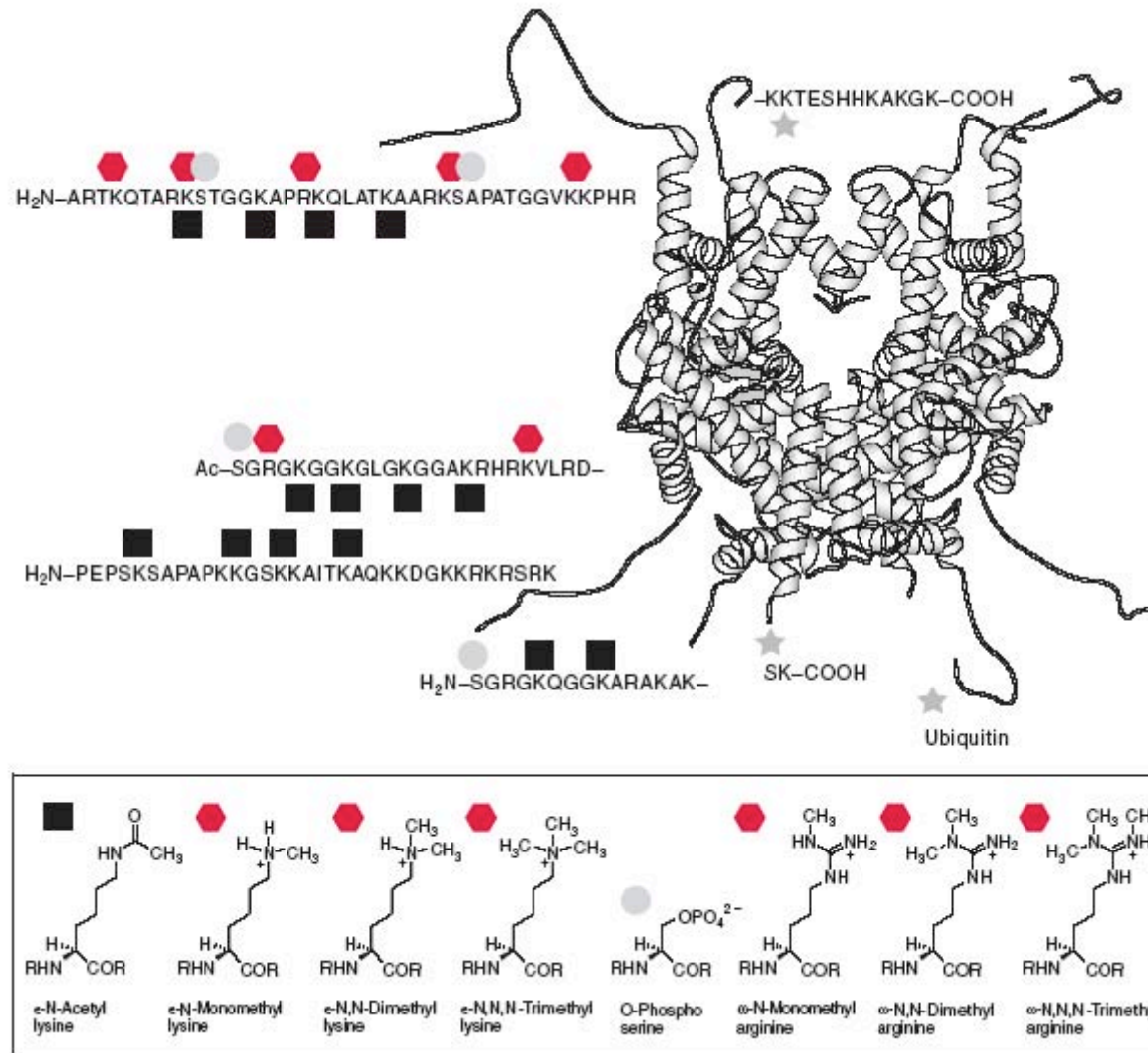
**Cytoplasm:**

- Cytoplasmic helices  $\alpha 5$ ,  $\alpha 6$ ,  $\alpha 7$ ,  $\alpha 8$ , and  $\alpha 9$  are labeled.
- CheW is shown bound to the C-terminus of the receptor.
- CheA histidine kinase is shown bound to CheW.
- CheB and CheC are shown bound to CheW.

# Protein O-Methylation

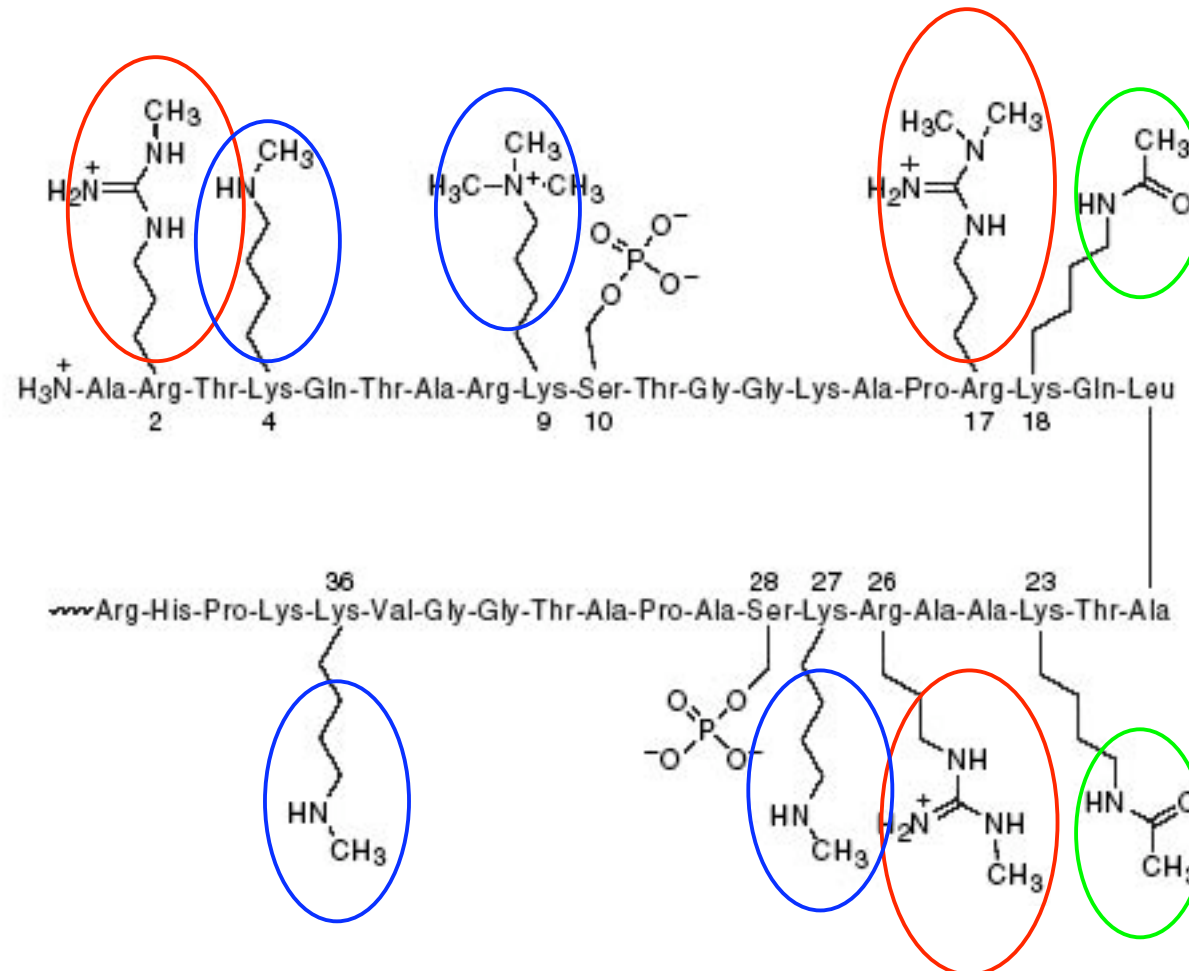


# Histone Methylation



## Function and Analysis of Post-translational Protein Modifications

# Histone Methylation in H3



Function and Analysis of Post-translational Protein Modifications

# Histone Modification

- H3 modification
  - 3 Rs methylated
  - 4 Ks methylated
  - 2 Ss phosphorylated
- Results in over 110,000 combinations of possible modifications
- approx.  $10^7$  possible nucleosomes



# Protein Phosphorylation

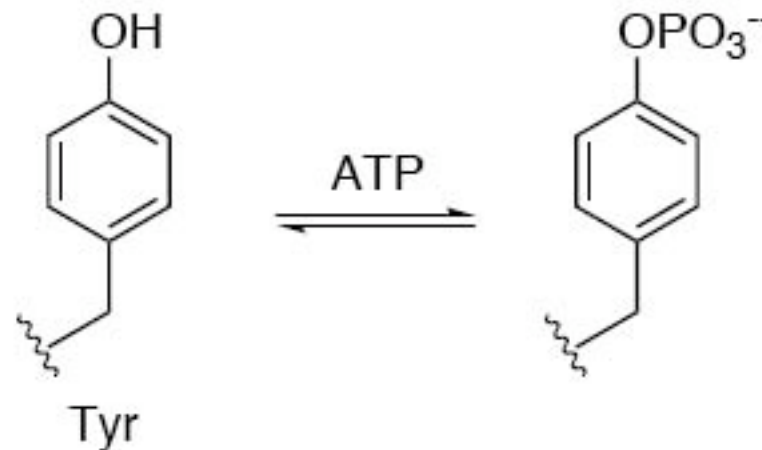
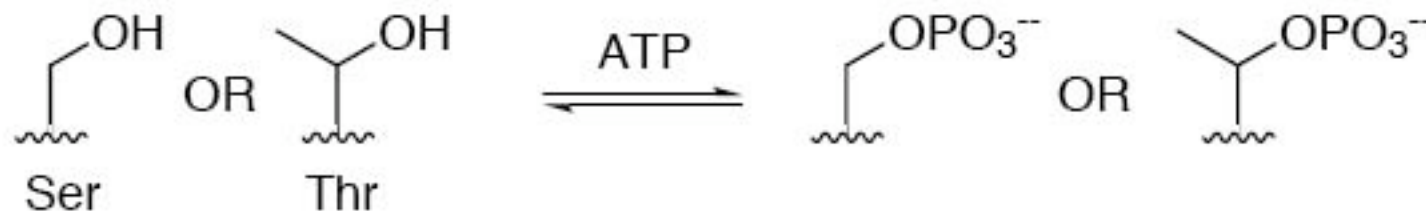
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# Protein Phosphorylation

- One of the most common PTMs
- Estimate that approx. 30% of all eukaryotic proteins become phosphorylated
- Human genome codes for approx. 2,000 protein kinases
- Transient
- Regulate a vast number of biological processes
  - Enzymatic activity
  - Metabolism
  - Motility
  - Signal transduction
  - Cell division
  - Cell growth
  - Apoptosis

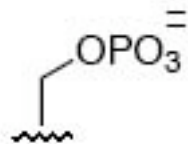
# Protein Phosphorylation

- Ser, Thr, Tyr phosphorylation by protein kinases

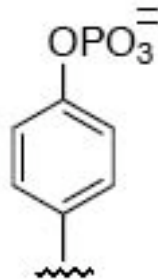


# Protein Phosphorylation

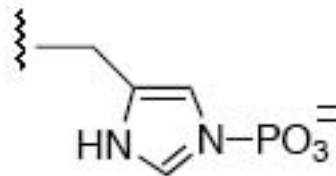
- Protein kinase variants defined by the type of protein side chain modification
  - a) Ser/Thr protein kinases (e.g. cAMP-dependent PK)



- b) Tyr protein kinases (e.g. insulin receptor kinase)

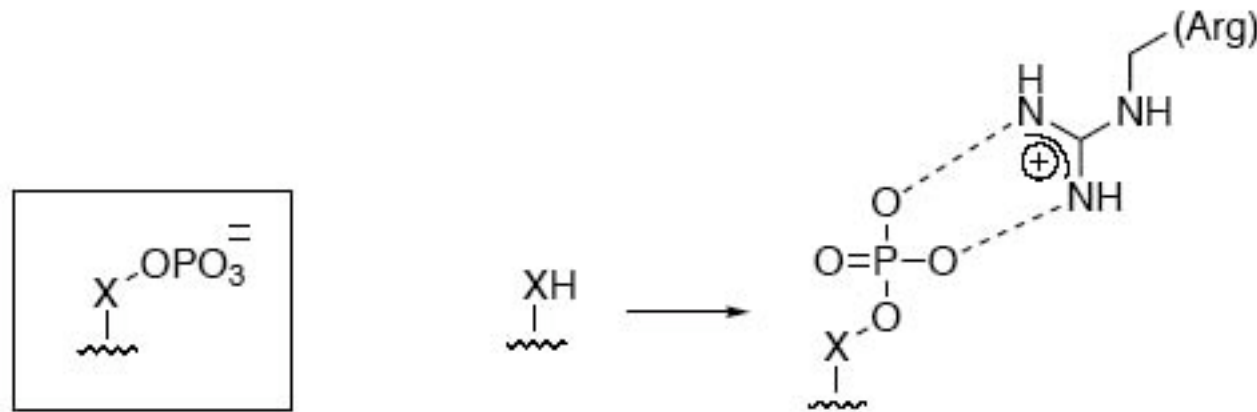


- c) His protein kinases (e.g. bacterial two component sensor/respons regulators)



# Protein Phosphorylation

- What is the purpose/effect of protein side chain phosphorylations?
  - Protein conformational switching
  - Introduction of a dianionic phosphate group ( $-PO_3^-$ ) induces electrostatic reorganization of local regions, loops of proteins
  - Reorganization often occurs via charge pairing with a cationic Arg side chain

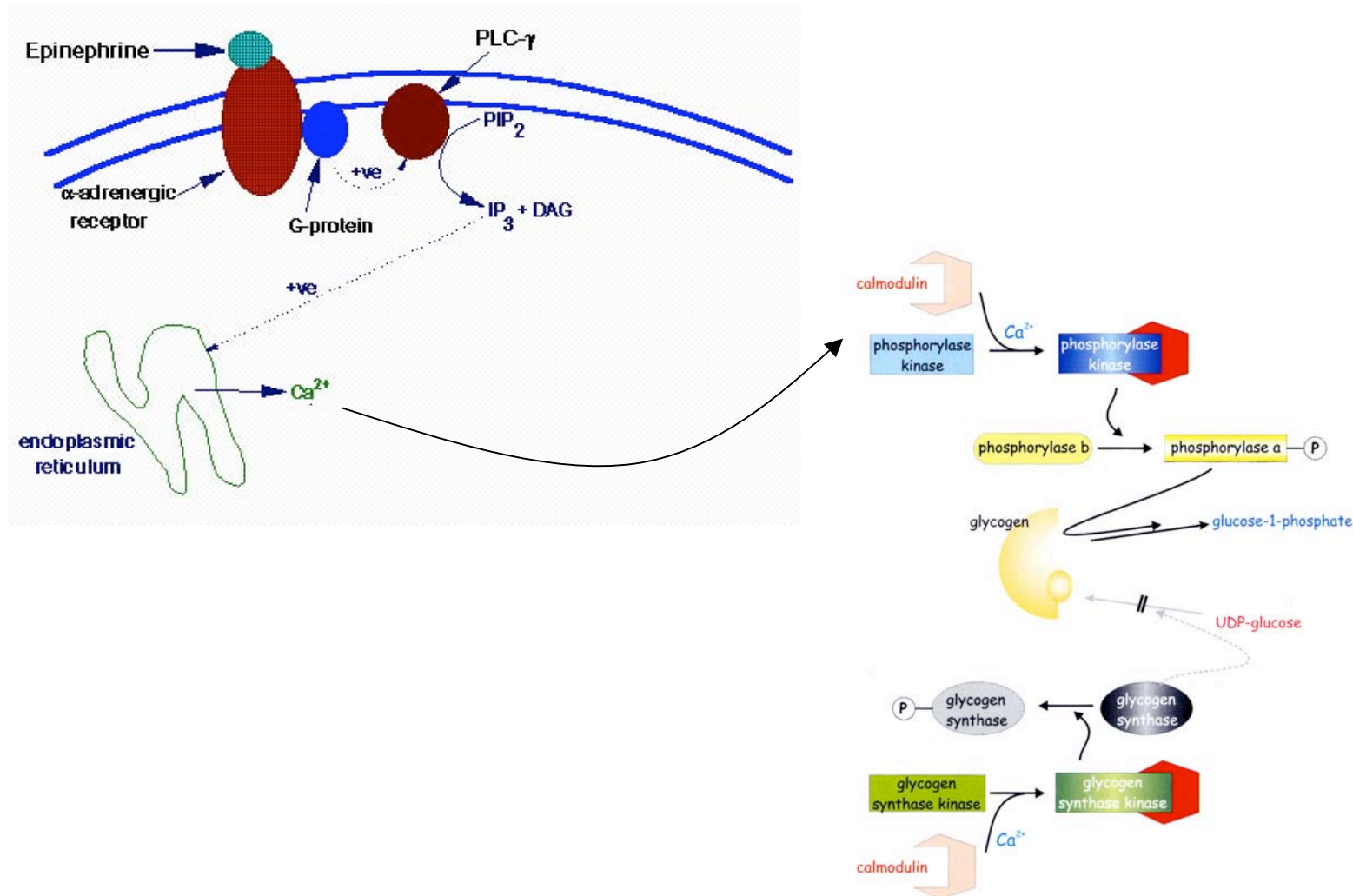


# Protein Phosphorylation

- What governs specificity of phosphorylation?
  - PKs recognise certain motifs

Protein Kinase	Recognition Motifs <sup>a</sup>	Phosphorylation Sites <sup>b</sup>	Protein Substrate (reference)
cAMP-dependent Protein Kinase (PKA, cAPK)	R-X- <b>S/T</b> <sup>c</sup> R-R/K-X- <b>S/T</b>	Y <sub>7</sub> LRRAS <b>L</b> AQLT F <sub>1</sub> RRL <b>S</b> IST A <sub>29</sub> GARRKA <b>S</b> GPP	pyruvate kinase (2) phosphorylase kinase, $\alpha$ chain (2) histone H1, bovine (2)
Casein Kinase I (CKI, CK-1)	S(P)-X-X- <b>S/T</b>	R <sub>4</sub> TLS(P)VS <b>S</b> LPGL D <sub>43</sub> IGS(P)ES(P) <b>T</b> EDQ	glycogen synthase, rabbit muscle (4) $\alpha_{s1}$ -casein (4)
Casein Kinase II (CKII, CK-2)	<b>S/T</b> -X-X-E	A <sub>72</sub> D <b>S</b> ESEDEED L <sub>37</sub> E <b>S</b> EEEGVPST E <sub>26</sub> D <b>S</b> ESEDEISNL	PKA regulatory subunit, R <sub>II</sub> (2) p34 <sup>cdc2</sup> , human (5) acetyl-CoA carboxylase (2)
Glycogen Synthase Kinase 3 (GSK-3)	<b>S</b> -X-X-X-S(P)	S <sub>641</sub> VPP <b>S</b> PSLS(P) <b>S</b> <sub>641</sub> VPPS(P)PSLS(P)	glycogen synthase, human (site 3b) (6,2) glycogen synthase, human (site 3a) (6,2)
Cdc2 Protein Kinase; CDK2-cyclin A	<b>S/T</b> -P-X-R/K <sup>c</sup>	P <sub>13</sub> AK <b>T</b> PVK H <sub>122</sub> STPPKKK <b>R</b> K	histone H1, calf thymus (2) large T antigen (2)
Calmodulin-dependent Protein Kinase II (CaMK II)	R-X-X- <b>S/T</b> R-X-X- <b>S/T</b> -V	N <sub>2</sub> YLRRRL <b>S</b> DSN K <sub>191</sub> MARV <b>F</b> SVLR	synapsin (site 1) (2) calcineurin (2)
Mitogen-activated Protein Kinase (Extracellular Signal-regulated Kinase) (MAPK, Erk)	p-X- <b>S/T</b> -p <sup>d</sup> X-X- <b>S/T</b> -P	P <sub>244</sub> <b>L</b> SP P <sub>92</sub> <b>S</b> SP V <sub>420</sub> <b>L</b> SP	c-Jun (7) cyclin B (7) Elk-1 (7)
Abl Tyrosine Kinase	I/V/L- <b>Y</b> -X-X-P/F <sup>e</sup>		

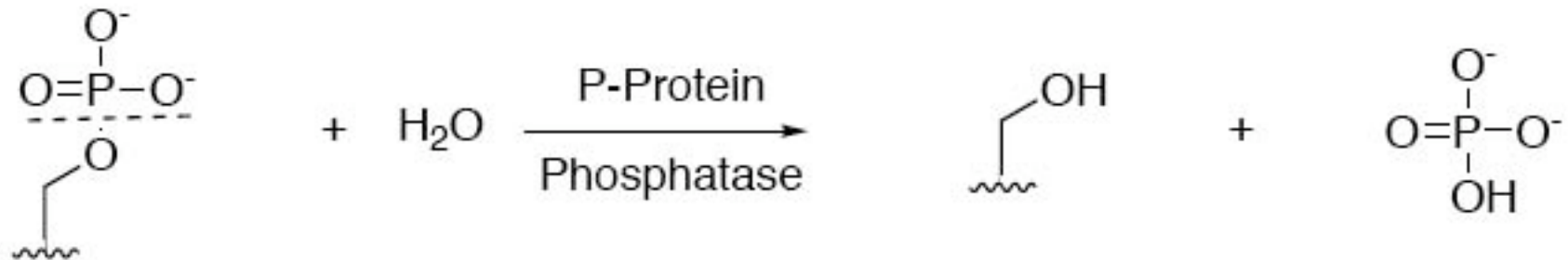
# Physiological Importance of Protein Phosphorylation



Function and Analysis of Post-translational Protein Modifications

# Protein Dephosphorylation

- Phosphoproteins are dephosphorylated by P-protein phosphatases
- Phosphatases counteract protein kinases
- Human genome encodes approx. 500 protein phosphatases
- Ser/Thr- and Tyr-specific phosphatases

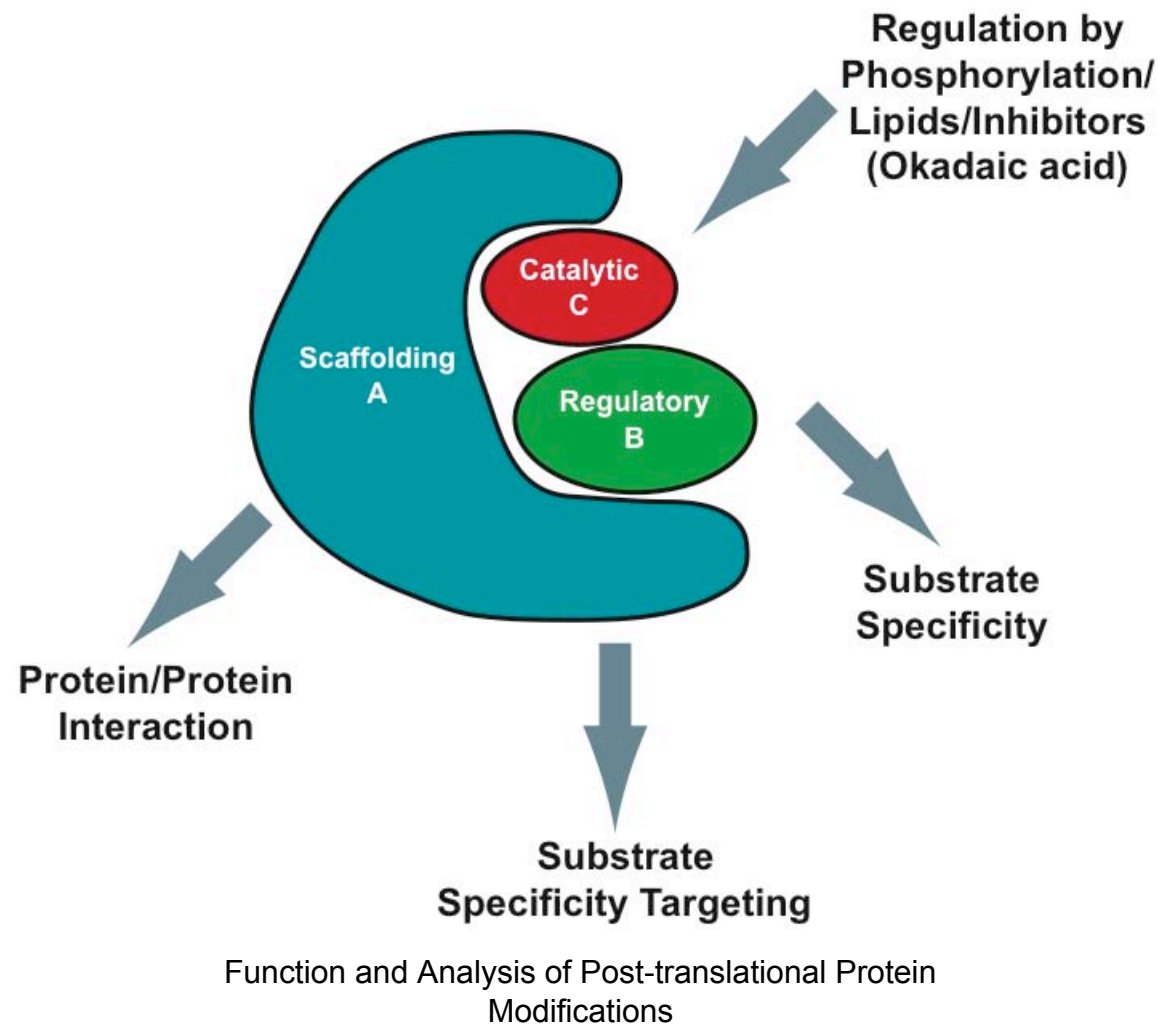




# Protein Phosphatases

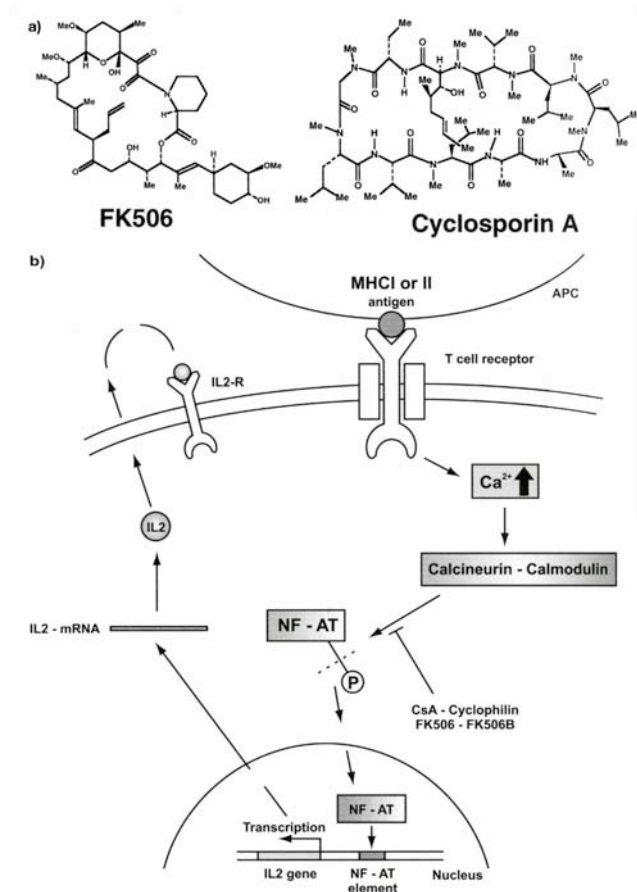
- Phosphatase 1
  - Ser/Thr-specific phosphatase
  - Regulates cell-cycle progression
  - Muscle contraction
  - Carbohydrate metabolism
- Protein Phosphatase 2A
  - Controls numerous cellular processes
  - Oligomeric enzymes

# Protein Phosphatase 2A



# Protein Phosphatase 2B, Calcineurin

- Ser/Thr phosphatase
- Controlled by cellular  $\text{Ca}^{2+}$
- Multimeric enzyme, binding to  $\text{Ca}^{2+}$ /calmodulin required for active enzyme

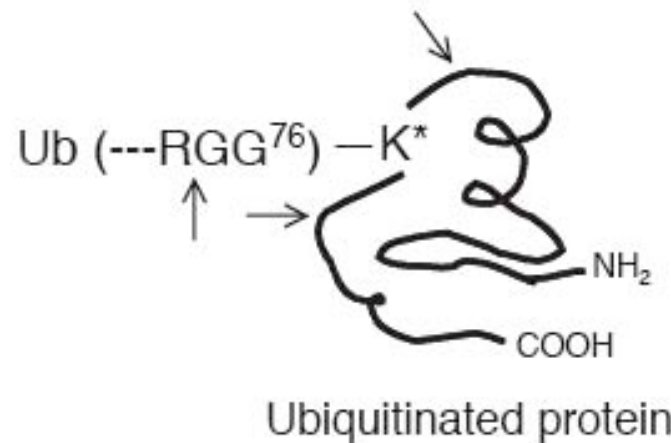


# Ubiquitination

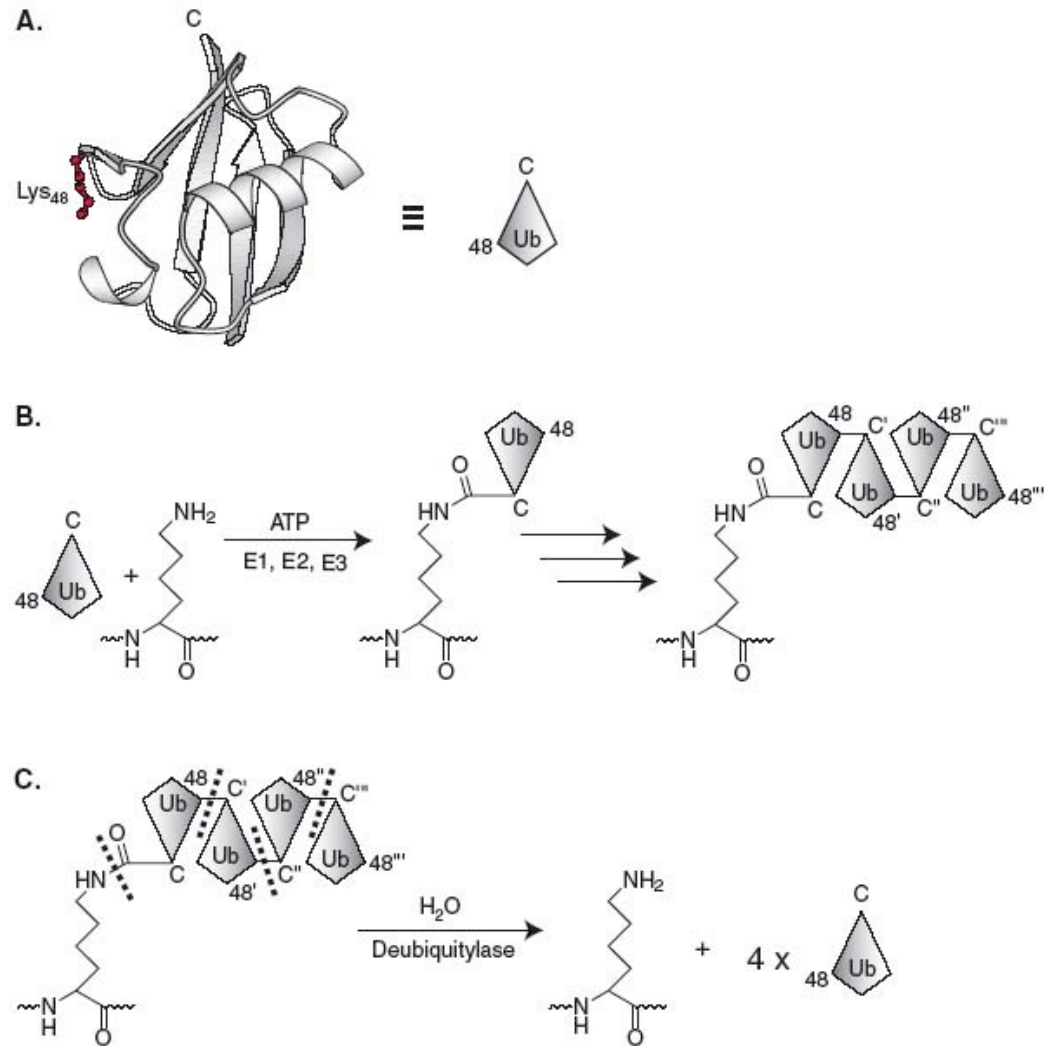
Function and Analysis of Post-translational Protein  
Modifications

# Ubiquitination

- Occurs in eukaryotes, not in prokaryotes
- Used for proteolytic destruction of specific proteins
- Ubiquitin is a small 76 residue protein
- Covalent attachment of multiple ubiquitin molecules to a protein substrate
- Degradation of the tagged protein by the 26S proteasome (ubiquitin is recycled)

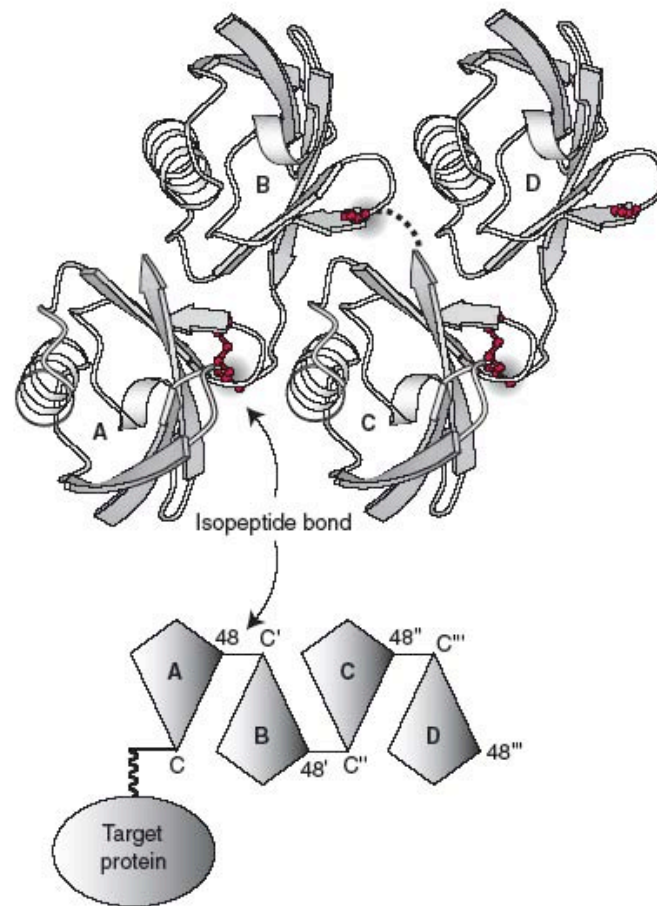


# Ubiquitination



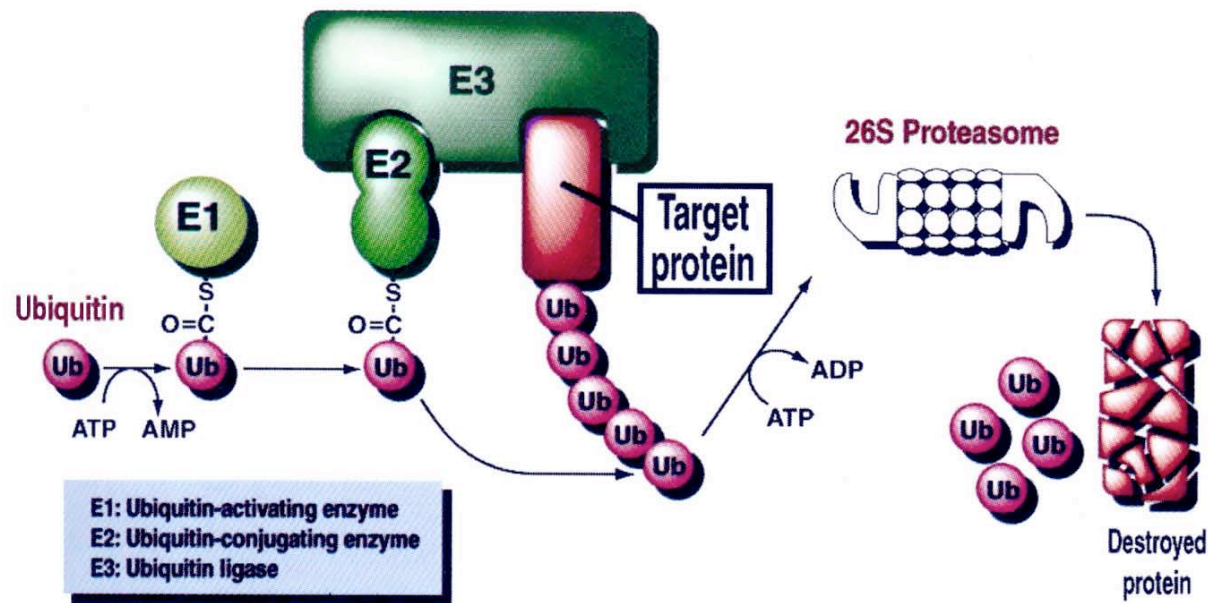
Function and Analysis of Post-translational Protein Modifications

# Ubiquitination



Function and Analysis of Post-translational Protein Modifications

# Ubiquitination Reaction Mechanism

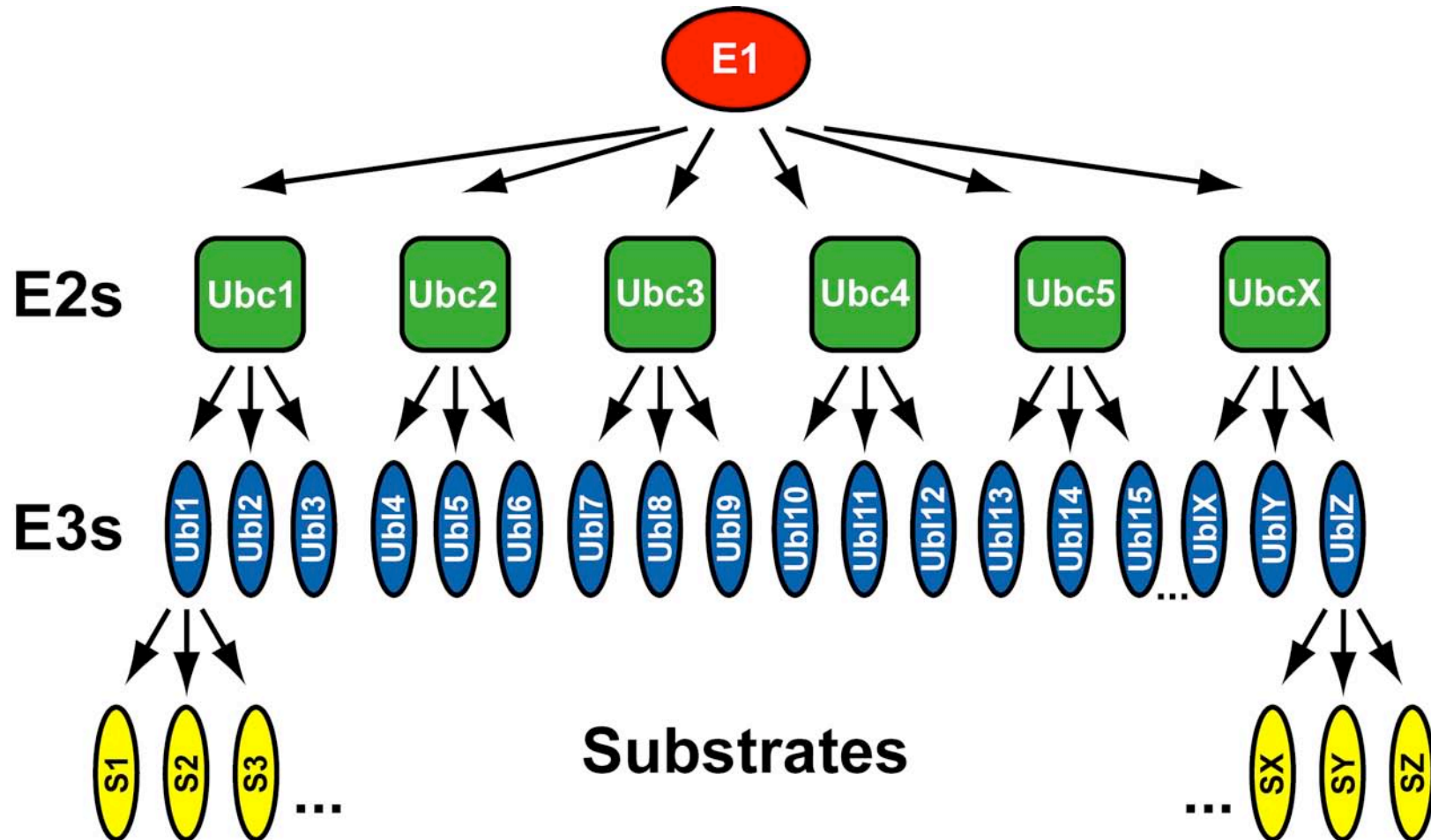




# Enzymes Involved in the Ubiquitination Process

- **E1:** only one known, consists of one or two polypeptidic chains, with M.W. 105 kDa per chain, very conservative protein
- **E2:** 5-12 proteins of this type known (depending on literature), homologous family
- **E3:** many and structurally unrelated, characterized by ultimate biological specificity, M.W. about 250 kDa
- **E4:** not known yet, but postulated as existing

# Ubiquitination



# SUMO (Small Ubiquitin-Related Modifier)

- SUMO does not have the Lys-48 found in ubiquitin
- SUMO does not make multi-chain forms
- SUMO-1,2,3 are the mammalian form
- SUMO-1: 101 amino acids, C-terminal Gly, 18% identical to ubiquitin



# SUMO Substrates

- Many of the known sumoylation substrates are nuclear proteins
  - p53
  - CREB
  - STAT1/4
  - GATA2, etc

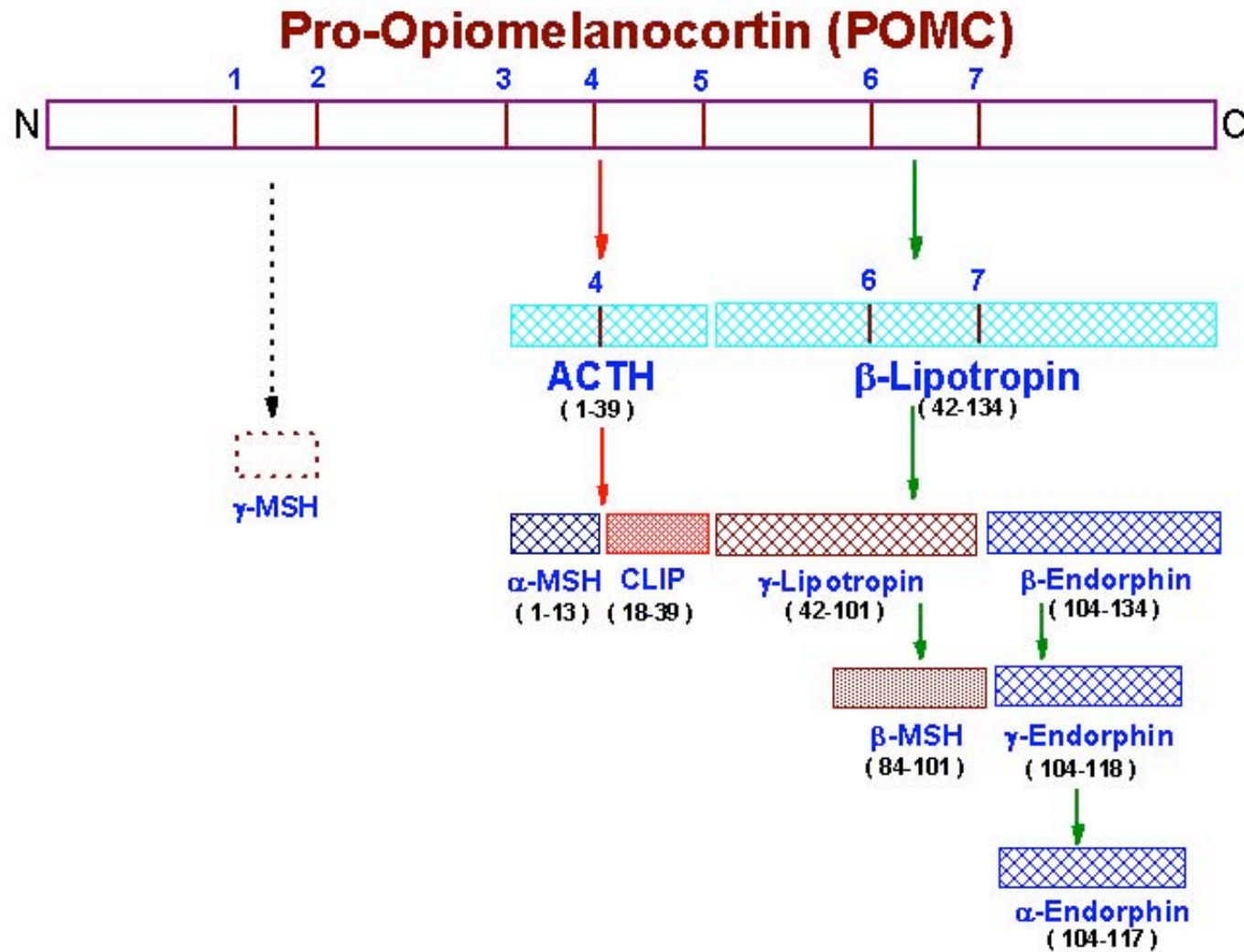
# Proteolytic Cleavage

Function and Analysis of Post-translational Protein  
Modifications

# Proteolytic Cleavage

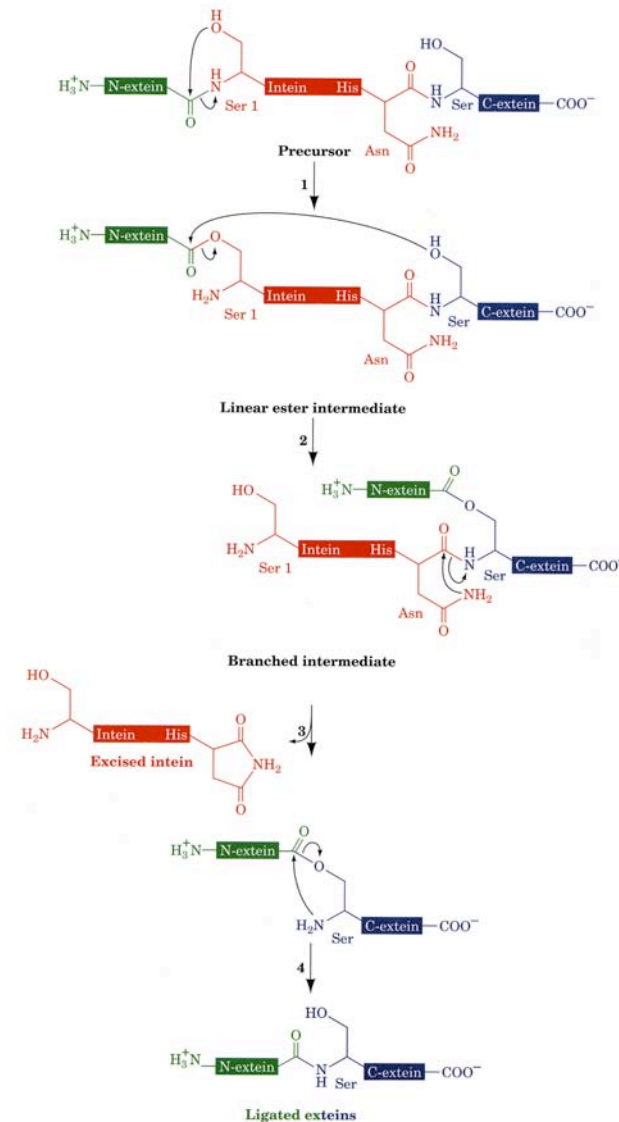
- Following translation, most proteins undergo proteolytic cleavage
- Removal of the initiation Met
- Many proteins are synthesised as inactive precursors
- E.g. pancreatic enzymes, enzymes involved in blood clotting = proproteins
- Activation occurs via removal of polypeptides

# Peptide Preprohormone



# Protein Splicing

- Internal protein sequences (intein)  
Excises itself from a surrounding
- External protein
- N- and C-terminal exteins are ligated
- Protein splicing occurs in bacteria and single-celled eukaryotes
- Exteins have no sequence similarity
- Inteins have conserved splice junctions:
  - Ser/Thr/Cys at the N-terminus
  - His-Asn/Gly dipeptide at the C-terminus
- inteins encode endonucleases that copy the intein gene into extein sequences
- Intein genes propagate themselves





# Glycoproteins

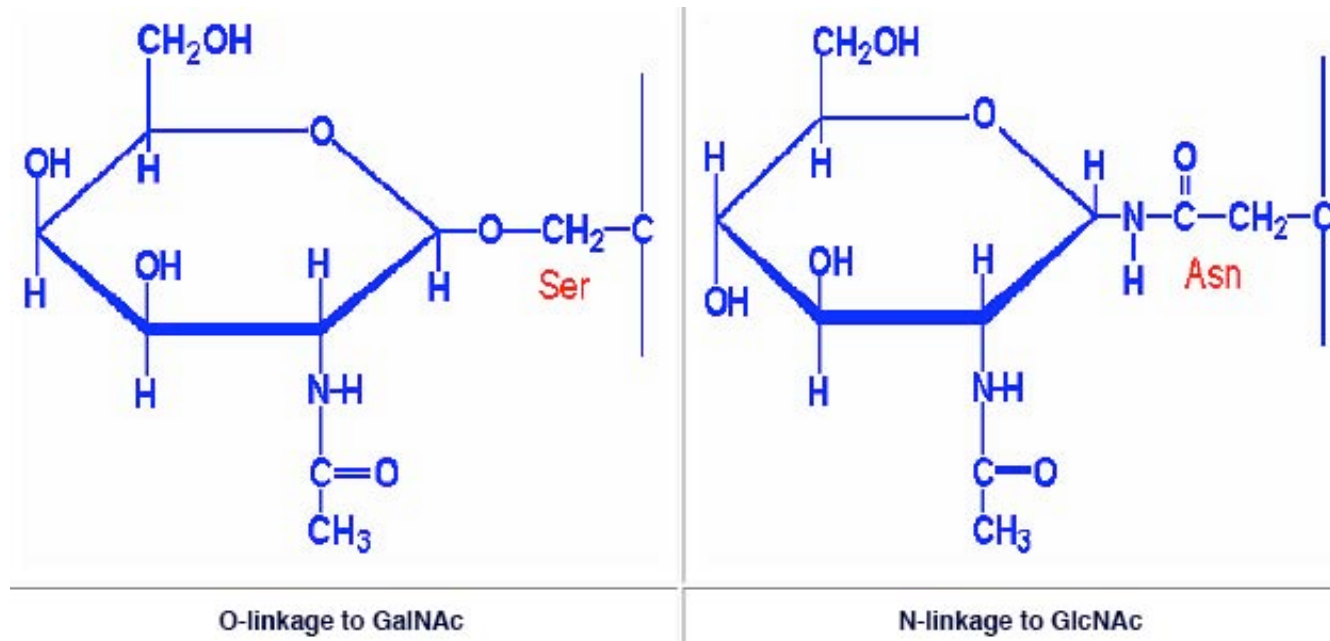
Function and Analysis of Post-translational Protein  
Modifications

# Glycoproteins

- Most secreted, or plasma membrane bound proteins are glycosylated
- Extracellular part modified
- Cytosolic and/or nuclear proteins also found to be glycosylated
- Predominant sugars found in glycoproteins are
  - Glucose, galactose, mannose, fucose, GalNAc, GlcNAc, NANA

# Glycoproteins

- Carbohydrates attached either O- or N-glycosidically
- N-glycosidic linkage through amide of Asn
  - Carbohydrate attachment within consensus sequence  
N-X-S(T)
- O-glycosidic linkage is through -OH of Ser, Thr, or OH-Lys

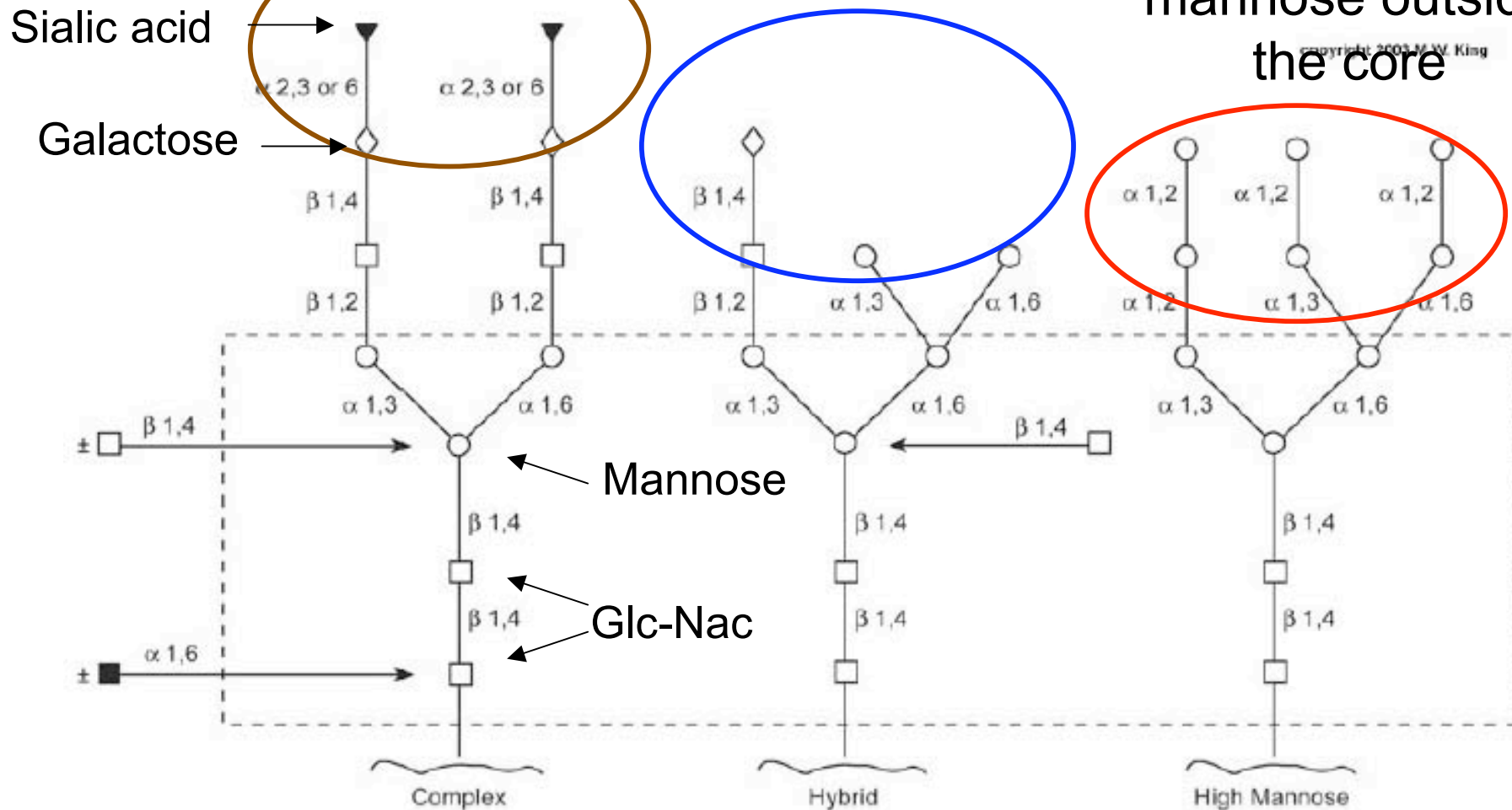


# Glycoproteins

Similar to hybrid,  
contains sialic acid

Contains various  
sugars

Contains only  
mannose outside  
the core

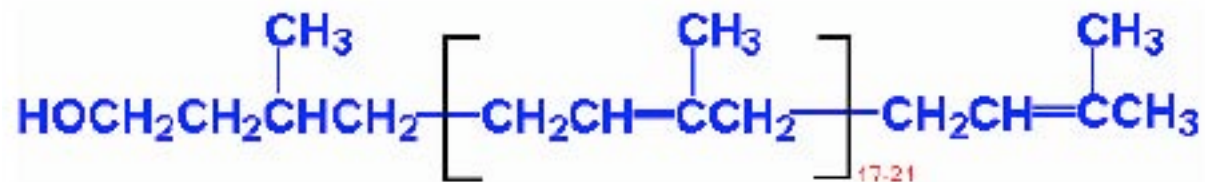


# Glycoproteins

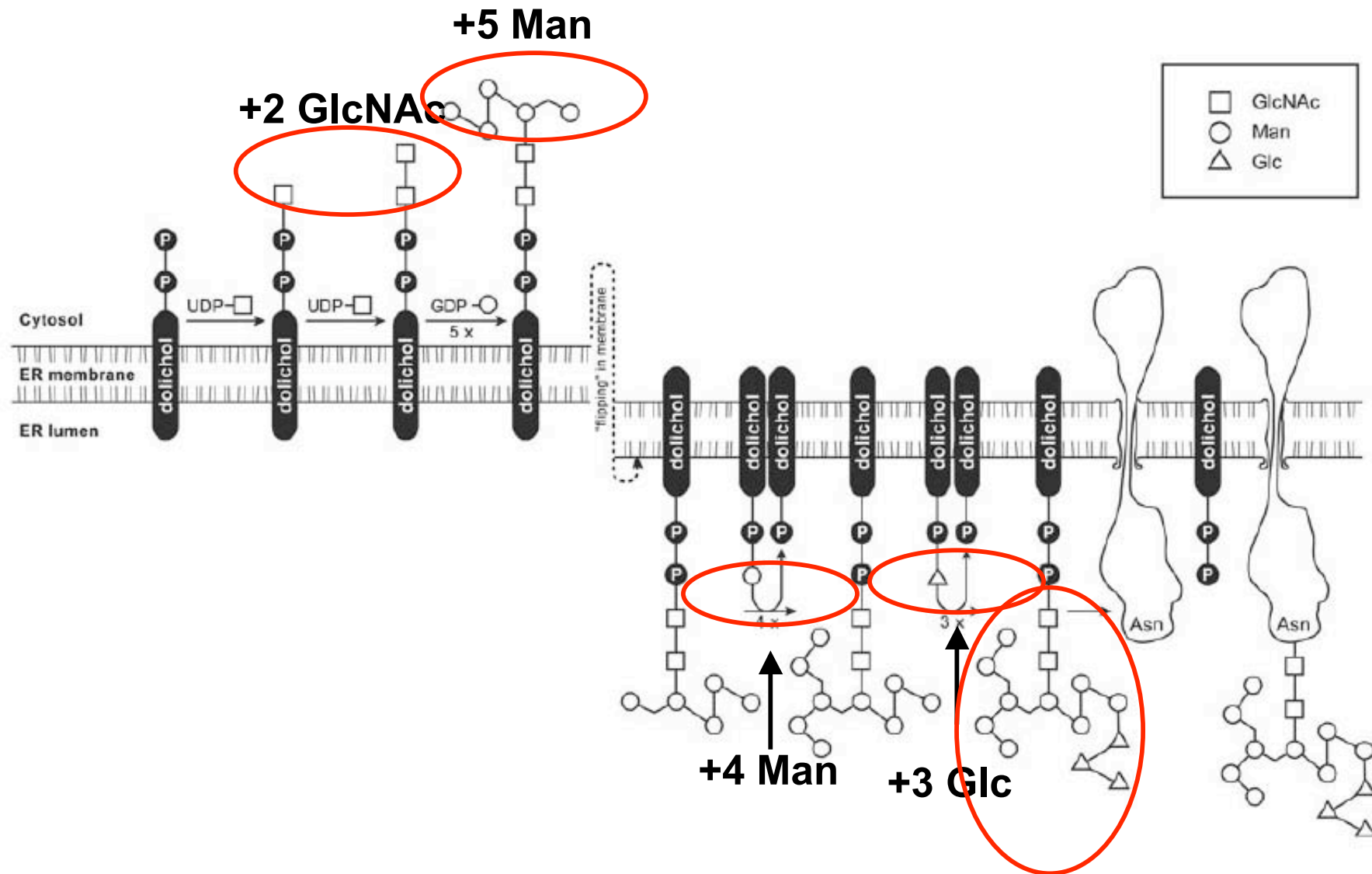
- Glycoproteins synthesised at rER
- Sugar attachment cotranslationally in the lumen of ER and continues in the Golgi for N-linked sugars
- O-linked sugars are attached post-translationally in the Golgi
- Sugars are activated by coupling to nucleotides
- Glc and GlcNAc are coupled to UDP
- Mannose is coupled to GDP

# Glycoprotein Synthesis

- N-linked glycoprotein synthesis requires the lipid intermediate dolichol phosphate

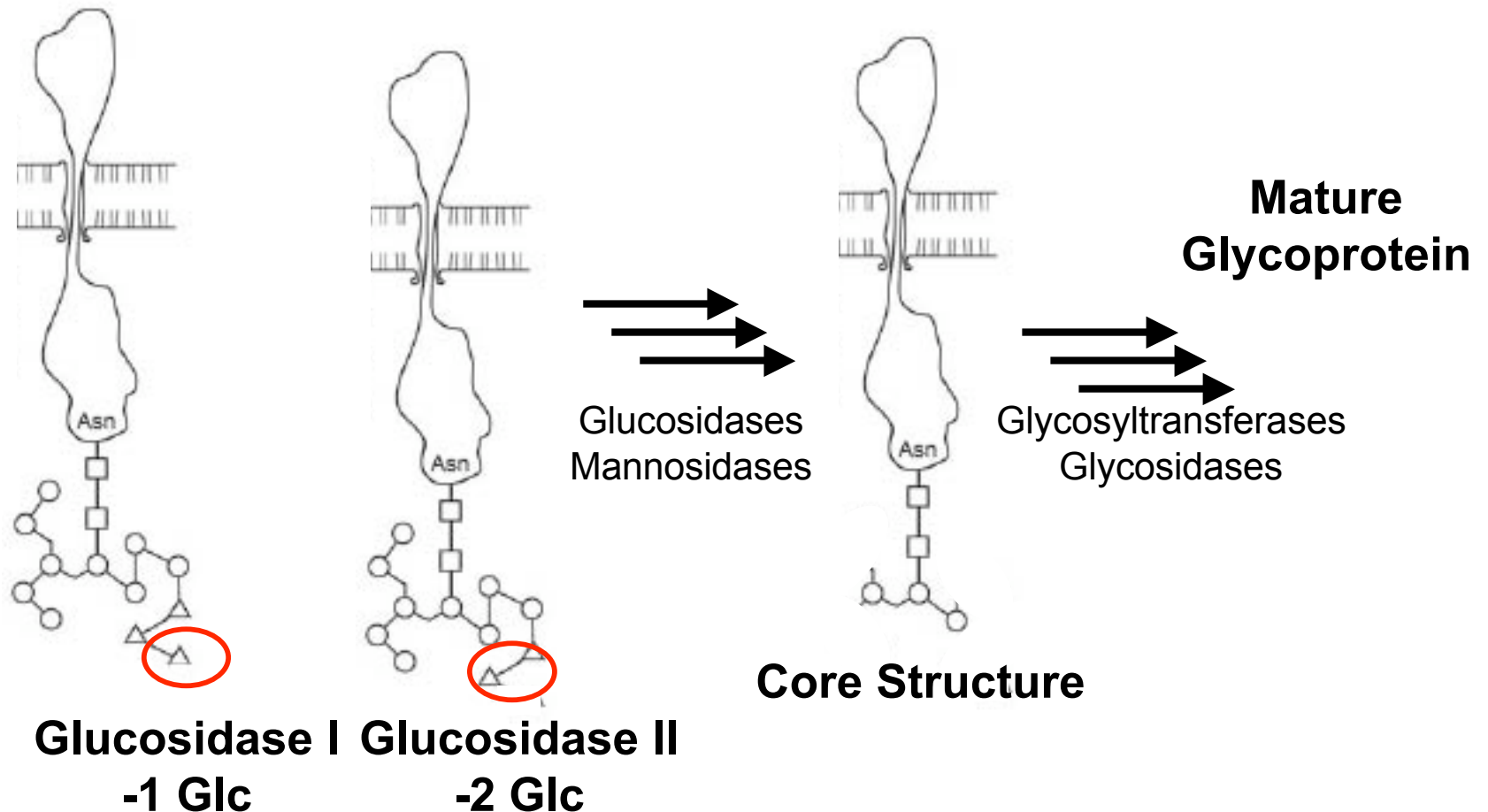


# Glycoprotein Synthesis



Function and Analysis of Post-translational Protein Modifications

# Sugar Trimming





# Clinical Significance of Glycoproteins

- ABO blood group antigens
  - ABO carbohydrates linked to lipids
  - ABO associated with proteins occur in the serum = secreted form
  - Some individuals produce secreted ABO
  - Used in forensic medicine
- Dystroglycan
  - Laminin receptor
  - alpha-dystroglycan serves as receptors for *Mycobacterium leprae* and other pathogens
- *Helicobacter pylori* attaches Lewis blood group antigen on the surface of gastric mucosa
- Etc. ...

## Further Reading

- Walsh, C.T. (2005) Posttranslational Modifications of Proteins. Expanding Nature's Inventory. Roberts and Company Publishers.
- Krishna, R. G. and F. Wold (1998). Posttranslational Modifications. Proteins - Analysis and Design. R. H. Angeletti. San Diego, Academic Press. 1: 121-206.
- Wold, F., (1981) In vivo chemical modification of proteins (post-translational modification) Ann. Rev. Biochem. 50,, 783-814.